PROCEEDINGS



AGRICULTURE 75 CONGRESS 2006



Agriculture For Life And Wealth Creation



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Preface

This is the book of Extended Abstracts of the Agriculture Congress on 'Agriculture For Life and Wealth Creation' held at Putrajaya, Malaysia from December 12 to 15, 2006. This Congress, the second, was organized by the Faculty of Agriculture, Universiti Putra Malaysia in collaboration with Ministry of Agriculture and Agro-based Industry (MOA), Ministry of Plantation Industries and Commodities (MPIC), Malaysia Agriculture Research and Development Institute (MARDI) International Society for Southeast Asian agricultural Sciences (ISSAAS) and The Incorporated Society of Planter (ISP).

The increasing demand on the agriculture sector to feed the continuous rising of would population coupled with limited resources available for agriculture has made agriculture a critical global issue. There is also a need to develop agriculture as a profitable and lucrative business to draw more people and investors into this sector. Based on these scenarios, the Faculty of Agriculture, Universiti Putra Malaysia took the initiative to organize Agriculture Congress 2006 to address and deliberate topics pertaining to the above theme. The congress covered various disciplines in agriculture, which include aquaculture, agricultural and animal production technologies, extension services, food processing and industries, food safety, genetic resources and utilization, molecular techniques in agriculture, microbial technology, marketing and policies, new crops, nutraceutical, agriculture pest management, plant nutrition and fertilizer, plantation management technology and sustainable agriculture.

A total of 157 papers were presented in 3 keynote, 12 plenary, 74 oral and 68 poster. The presentations from 10 countries indicate the important of addressing these issues to ensure the agriculture sector is attractive and profitable.

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> Dzolkhifli Omar Lau Wei Hong Jothi Malar Panandam Mohamed Hanafi Musa Mohd. Salleh Kamarudin Md. Ariff Hussein Mohd. Razi Ismail Tee Tuan Poy

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SCREENING AND SELECTION OF ACID SOIL TOLERANCE INBRED LINES IN MAIZE (ZEA MAYSL.)

PKD Hayati¹, GB Salch¹, J Shamshuddin² and S Napis³

Department of Corp Science, Department of Land Management, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, Faculty of Agriculture, UPM," Department of Cell and Molecular Biology, Faculty of Agriculture, UPM, "Department of Cell and Molecular Biology, and Biology

INTRODUCTION

Malaysia has been very much dependent on the importation of grain maize from other countries in the past adecades, while local production is negligible (FAOSTAT, 2005). In order to achieve sufficiency, it is necessary develop the local maize growing industry to reduce the dependence on imported maize in the long-term. However, most of the anable soils in the country are acklic. The highly weathered soils which are classified as Utisote and Oxisots occupy 72 % of land area in Malaysia (IBSRAM, 1985). The soils have low cation exchange capacity (CEC), high soil solution aluminum concentration and are usually deficient in Ca and/or Mg. Some of these soils being planted with com during the early years of rubber and oil pulm replanting phase, but the productivity is in these soils due to Al toxicity and Ca and/or Mg deficiencies (Shamshuddin et al., 1991). These soils can be highly productive by application of lime (Shamshuddin et al., 1989; Ismail et al., 1993). However, the application of lime in many tropical countries may not economical and permanent solution.

Planting acid soil tolerant maize varieties offer the strategy for improving maize production under acidic soil identification of acid soil tolerant and high yielding inbred lines is potential to produce hybrids with complementary performance. Field trials have been proven to be effective in selecting aluminum tolerant plants, but are expensive and time consuming. A rapid screening method is needed to identify and select a large number genotypes in plant breeding. Screening using hernatoxylin staining of seedling root has been widely use screening relatively large populations based on the formation of an intense blue coloration in the root tips of seasoning relatively large populations based on the formation of an intense blue coloration in the root tips of seasoning relatively large populations based on the formation of an intense blue coloration in the root tips of seasoning relatively large populations based on the formation of an intense blue coloration in the root tips of seasoning relatively large populations of the root can be assessed (Polle et al. 1978; Ownby, 1993).

Laboratory- and greenhouse-based techniques have been developed to identify Al-tolerant and Al-sensitive precisely, i.e. using nutrient solution culture and pot trials. Since the main effect of Al toxicity is the inhibition agrowth, the roots are not easily observed using soil culture. Screening in nutrient solution allows studying Alexandrope growth and perfect of the providing easy access to the root system and strict control over nutrient availability and pH (Blamey et al. Some researchers use this technique to screen maize genotypes that were tolerant to Al (Kasim and Wassen, Cancado et al., 1997; Giaveno and Minmuch Filho, 2000). However, many nutrient solution culture studies with high ionic strength of nutrient solution with high Al concentrations. In low nutrient solution concentrations that result in decreased plant performance have been similar to those reports.

Screening techniques using soil in glasshouse are very representative of real conditions in the field (However, Soil bioassays seem to reproduce more realistic field condition (Giaveno and Miranda Filho, 2002). In the use of hematoxylin staining assay and screening in nutrient solution using low ionic strength concentration of Al in maize is scarce. The objective of this study was to screen and select maize inbred the to aluminum toxicity using hematoxylin staining assay, screening in nutrient solution and screening in portrain.

METHODOLOGY

Genotype. Thirty-six genotypes consisted of 6 maize inbred lines from CYMMIT, 2 composites, 1 hours locally-developed inbred lines were used in hematoxylin staining assay and screening in nutrient solution. In the maize inbred lines short listed from these experiments were further evaluated in pot experiment.

NBRED LINES IN MAIZE (ZEA

S Napis

partment of Cell and Molecular Biology, Faculty at

ce from other countries in the past was achieve sufficiency, it is necessar a corted maize in the long-term. However, it is necessar as order maize in the long-term. However, in the analysis of these seasons places, but the productivity is long phase, but the productivity is long phase.

naize production under acidic sector produce hybrids with complete administration tolerant plants, but a dentify and select a large number of earlier proof has been widely section in the root tips of sector forms a complex with Al so the Ownby, 1993).

entify Al-tolerant and Al-sensities of Al toxicity is the inhibition river solution allows studying A mailability and plot (Blumey at a bleam to Al (Kasim and Wassemutrient solution culture studies and the low mutrient solution concerns to be been similar to those reports.

d conditions in the field (Hoselend Miranda Filho, 2002). In the film using low ionic strength and select maize inbred the solution and screening in passes.

YMMIT, 2 composites, 1 = mand and screening in nutrient solution. Hematoxylin staining assay (HS). Maize seeds were surface sterilized with 1.5% and germinated inside the moisture paper until the roots were 2 cm or more in length. Three aluminum concentrations (5, 10 and 20 μM) were seed to screen genotypes tolerant to Al toxicity in nutrient solution. The basic protocol was based on Polle et al. = 78). The roots of seedlings cultivated for 17 hours were gently shaken with distilled water for 30 minutes. The water was then replaced by 0.2% hematoxylin solution (2g hematoxylin +0.2g KI) for 15 minutes and was replaced as the distilled water for 30 minutes. After staining, the photograph was taken and the pattern of HA was used to take all genotypes into tolerant, moderately tolerant, moderately sensitive and sensitive based on no or less stain at complete staining of the root tips.

in nutrient solution (NS). Initially, twelve genotypes and 4 Al levels (0,10,20 and 30 μM) were used to the Al concentration that could give different response among Al-tolerant and Al-sensitive based on the root and aluminum tolerant index variables under the absence and the presence of Al in aerated and low ionic NS (1332.74 μM). Al was added to the NS from 0.1M stock of AlCl_b.6H2O and the pH was adjusted to daily. The root growth variables were net seminal root length (NSRL), ratio seminal root length (RSRL) weight (RDW) and shoot dry weight (SDW). Aluminum tolerant index was based on the ratio of the root and without Al.

in pot trial (PT). Pot experiment was conducted using two aluminum levels, i.e. acid soils with and ime. GML was applied at 2 t ha⁴, 30 days before planting. The plant height and plant top dry weight were a tays after planting. The chemical properties of the soil are Table 1.

the chemical properties of the soil

	pH(H ₂ O)	K	Na	Ca	Mg	CEC	Al	AI saturation	SOM
we widity	1:1	1:1	cmol _c kg-1				%	%	
wede Soil	4.6	0.06	0.21	1.41	0.1	6.67	1.82	51	1.49
War Acidic	5.1	0.13	0.38	2.94	0.29	7.86	0.34	27	

Design. Screening in NS and PT was conclucted in a split plot design with 3 replications. Main plots meentrations or soil acidity and sub plots were genotypes. Analysis of variance was calculated for each means of 6 sec

RESULTS AND DISCUSSION

staining assay (HS). There was no root stain in 5 μM AI for all genotypes, indicating there was no secreted highly sensitive to AI. However, there was a high variation of root staining pattern in 10 and 20 μm and the intensity of stain in the root tip in 20 μM, while the AI-sensitive genotypes rice stain in 10 μM and the intensity of stain in the root increased in 20 μM AI. A visual identification are revealed HS assay was sufficient to discriminate between AI-tokerant and AI-sensitive genotypes.

3. SM7-6, SM7-10, SM7-11, SM7-12, IPB-12, CML-1 and CML-6 were characterized as AI-sensitive, CML-3, IPB-18, SM5-4, and SM5-5 as AI-sensitive. Other genotypes were moderately tolerant and moderately sensitive to AI toxicity.

matrient solution (NS). Table 2 shows highly significant effects of aluminum concentrations and
 mot growth variables. The interaction of aluminum and genotype also shows highly significant for
 mot growth variables.

Table 2. Mean squares for responses of NSRL, RSRL, RDW and R/S of 12 genotypes

Source of			Me	an squares			
Variation	D.F.	NSRL	RSRL	RDW	R/S		
Replicate	2	0.89	0.04	0.000002	0.0001		
	3	1110.30**	10.37**	0.000122**	0.0251**		
Aluminum			0.01	0.000001	0.0005		
Error (a)	6	0.21	T	0.000055**	0.1977**		
Genotype	11	23.63**	0.18**		0.0015		
AlxGen	33	7.92**	0.08**	0.000004**			
Error (b)	88	1.50	0.03	0.000002	0.0009		

^{**, *} significant at p<0.01 and p<0.05, respectively

Analysis of variance based on the aluminum tolerant index variables was consistent with root growth However, only Relative NSRL showed highly significant for the interaction of aluminum and generate results are similar to Kasim and Wassom (1990) that used 18 maize genotypes at 83, 111 and 148 μM agrowth decreased significantly as Al concentration in solution increased (Fig. 1). Al affected root growth decreased significantly as Al concentration in solution increased (Fig. 1). Al affected root growth since roots absorb Al from the solution. The most common symptom of Al toxicity is a stantal root spitch, stubby and show little branching. The interaction of aluminum and genotypes was significant in RSRL, RDW, and Relative NSRL. Hence, the appropriate comparison of mean would be between generate aluminum concentration (Table 3). All variables showed variation of tolerant to Al among the same aluminum concentration among 12 genotypes at 30 μM Al on Rel-NSRL. The highest mean appropriate to the same position at 30 μM Al. The concentration of 20 μM Al was sufficient to greater the properties along genotypes based on Al tolerant index.

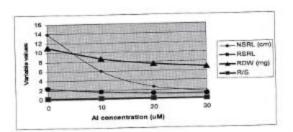


Fig. 1. Effect of 4 Al concentrations on root growth for 12 genotypes

Table 3, Mean squares for responses of Relative NSRL, NSRL, RSRL and RDW at 10, 20 and 30 µM &

Altoria	Relative NSRL	NSRL	RSRL	RDW
Al(uM)	0.11**	21.15**	0.16**	0.00002**
0	0.14**	22.48**	0.06**	0.00001**
30	0.096	3.39**	0.02*	0.00001**

^{**, *} significant at p=0.01 and p=0.05, respectively

Using 36 genotypes and 2 Al concentrations (0 and 20 µM Al), aluminum and genotype showed effects on all root growth variables. However, the interaction of aluminum and genotype was high selections of aluminum and genotype was high selections.

R/S of 12 genotypes

ean squares		
RDW	R/S	
0.000002	0.0001	
0.000122**	0.0251**	
0.000001	0.0005	
0.000055**	0.1977**	
0.000004**	0.0015	
0.000002	0.0009	

coles was consistent with root grown interaction of alternium and genor as a genotypes at 83, 111 and 148 µM rassed (Fig. 1). All affected root grown of Al toxicity is a sturied root grown and genotypes was significant in son of mean would be between grown arisino of tolerant to Al among the lon Rel-NSRL. The highest mean ground and on 620 µM Al was sufficient to the state of the

Land RDW at 10, 20 and 30 gate #

SW (me)

RSRL	RDW
0.16**	0.00000
0.06**	0.00000
0.02*	0.00000=

inum and genotype showed him and genotype was him a

Mean squares for responses of NSRL, RSRL, RDW and R/S of 36 genotypes

DO STEAM		-	M	ean squares	
ETHER	D.F	NSRL	RSRL	RDW	R/S
micate.	2	1.4	0.8	0.00001	0.02
Bericum	1	7731.9**	138.7**	0.0031**	2.16**
(a)	2	1.1*	1.4	0.00001**	0.02
mun pe	35	22.3**	1.9**	0.0002**	0.77**
Mim	35	14.3**	1.5	0.00017	0.56**
200	146	2.5	0.3	0.00003	0.01

maificant at p≤0.01 and p≤0.05, respectively

the marking of means in each root growth and aluminum tolerant index variables, SMRG, IPB-14, SM7-6, and IPB-12 were classified as Al-tolerant, CML-5, IPB-8, IPB-11, IPB-15, IPB-17, IPB-18, IPB-19, SM5-5-6-A, Sel-G and Suwan as Al-sensitive and another genotypes as moderately tolerant and moderately aluminum toxicity.

n pot trial (PT). Analysis of variance for plant height and plant top dry weight showed significant for soil

significant for genotype. However, there was no interaction between soil acidity and genotypes.

significant for genotype. However, there was no interaction between soil acidity and genotypes.

significant for genotypes. IPB-13, IPB-15, IPB-20, SW-2,

12, Putra J-58 and Sel-G were classified as Al-sensitive genotypes. IPB-8 as Al-sensitive inbred line

significant result on 2 series pot experiments, while SMRG, SM7-11, IPB-12, IPB-14, CML-1, SM7-6,

SM 5-5, SM 5-4, SM 5-9, IPB-19, MI-13, Suvara, Sel-A, CML-3 and IPB-18 were classified as

anotypes due to no significant differences for plant height and plant top dry weight under acidic and non

CONCLUSION

acoxylin staining assay, screening in nutrient solution and screening in pot trial, 36 maize genotypes field into four groups i.e. tolerant, moderately tolerant, moderately sensitive and sensitive to Al. The Allines are IPB-12, SM 7-6 and SM 7-11 and the Al-sensitive inbred lines are IPB-8 and IPB-11.

REFERENCES

- C., D.C. Edmeades, C.J. Asher, D.G. Edwards and D.M. Wheeler. 1991. Evaluation of solution culture for studying aluminum toxicity in plants. In R.J. Wright et al. (Eds.) Plant-soil Interactions at Low pH,
- EMA, L.I. Loguercio, P.R. Martins, S.N. Parentoni, E. Paiva, A. Borem and M.A. Lopes. 1997.
 Emany in staining as phenotypic index for aluminum tolerance selection in tropical maize (Zea mays L.).
 - 2005, World Development Indicators, ESSA, 3p.
- D, and J.B. Miranda Filho. 2000. Selection methods for maize seedlings in greenhouse as related to tolerance. Scientia Agricola 59/41:807-810.
- D., and J.B. Miranda Filho. 2002. Field comparison between selection methods at the maize seedling sciation to aluminum tolerance. Scientia Agricola 59(2):397-401.
- R.H. 1991. Identifying plants adaptable to low pH conditions. In R.J. Wright et al. (Eds.) Plant-soil actions at Low pH, 885-904
- 1985. Report of the Inaugural Workshop and Proposal for Implementation of the Acid Tropical Soils security Network. International Board of Soil Research and Management. Bangkok. 40p.
- Shamshuddin and S.R. Syed Omar. 1993, Alleviation of soil acidity in Ultisol and Oxisol for com-Pant and Soil 151:55-65.

- Kasim, F., and C.E. Wassom. 1990. Genotypic response of com to aluminum stress. I. Seedling test for measure aluminum tolerance in nutrient solutions. Indonesian J. of Crop Sci. 5(2):41-51.
- Ownby, J.D. 1993. Mechanism of reaction of hematoxylin with aluminum treated wheat roots. Phys. 37:371-380.
- Polle, E., C.F. Konzak and J.A. Kittrick. 1978. Visual detection of aluminum tolerance level in byhematoxylin staining of seedling roots. Crop Sci. 18:823-827.
- Shamshuddin, J., H.A.H. Sharifuddin and Z. Shamsuddin. 1989. Reduction of aluminum toxicity through UPM Research News. 3(2): 2-3.
- Sharnshuddin, J., I. Che Fauziah and H.A.H. Sharifuddin. 1991. Effects of limestone and gypsum accidental Malaysia ultisol on soil solution composition and yields of maize and groundnut. Plant and Soil 134.