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The Effect of Supplementation of Phytase on Broiler Rations to the Retention of Phosphor, Calcium and Nitrogen

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Abstract: The research was conducted to determine the optimum dose of phytase supplementation of *Fusarium verticillioides* on Phosphor-deficient rations. This study used 24 broiler chickens (4 weeks). This research used Completely Randomized Design (CRD) with five ration treatments with 4 replications. Ration treatment was supplemented phytase with different doses: R1: 0 U/kg, R2: 250 U/kg, R3: 500 U/kg, R4: 750 U/kg and R5: 1000 U/kg. Ration was based on iso protein (20%) and iso energy (2882 kcal/kg). The variables measured were retention of phosphor, calcium and nitrogen. The results showed that supplementation of phytase on broiler rations which Phosphor-deficiency was influenced the retention of Phosphor, calcium and nitrogen significantly ($P < 0.01$). The optimum phytase supplementation of *Fusarium verticillioides* on broiler rations which phosphor-deficient was 750 U/kg ratio, it is seen from retention of phosphor (71.38%), calcium (75.65%) and nitrogen (67.61%).

Key words: Phytase, supplementation, *Fusarium verticillioides*, retention, broiler

INTRODUCTION

Feed is the major factor that determines the success of the animal farm business. On managing poultry farm intensively feed should always be available and adequate. The main feed ingredients of poultry ration formulation largely derived from plant sources (80-85%) for the source of energy, protein and minerals, especially phosphor minerals. The availability of phosphor in plant as a raw material of monogastric livestock feed mainly poultry is limited by the presence of phytic acid anti-nutrients that binds P-elements and other elements, hence, they can not be utilized. Phosphor that is bound to phytic acid can not be digested and absorbed by monogastric livestock, especially poultry. Phytase was not produced in the digestive tract. The unavailability of phytase in digestive tract of poultry, will make phosphor be excreted with the excreta (Mallin, 2000).

In the normal physiological conditions phytic acid will bind essential minerals such as Ca, Mg, Fe and Zn. Beside it can bind the amino acid and protein and inhibit digestion by digestive enzymes (Pallauf and Rimbach, 1996) which to be lower of availability and digestive power, so that phytic acid is considered as anti-nutrients in the feed. Hence, an enzyme was given to hydrolyse it. One of the enzymes that can decrease phytic acid anti-nutrients is by supplementing phytase on the ration.

Phytase can hydrolyze phytic acid to inorganic phosphor and myo-inositol phosphate derivate, thus increasing the availability of Phosphor, digestibility and utilization of feed protein and amino acids. Phytase enzyme is widespread in nature because it can be found in

microorganisms (Shieh and Ware, 1968), so it can to be applied in the feed. Some of the phytase of microorganisms have been characterized as phytase from *Escherichia coli*, *Bacillus subtilis* and *Aspergillus*. Supplementation of phytase can increase the availability of phosphor and calcium. Phytase supplementation in broiler rations which low phosphor can increase the availability of nutrients, weight gain and feed conversion and increase phosphor and calcium retention significantly (Mondal *et al.*, 2007). Further, addition of 1000 U phytase/kg in broiler rations is capable to increasing the availability of Arginine 3.7% and Nitrogen 4.2% and Calcium 16.6% (Lan *et al.*, 2002).

In this study the phytase produced by *Fusarium verticillioides* Phosphor is expected to hydrolyze phosphor that bound in phytic acid without losing its activity. Yetti *et al.* (2010) reported, the phytase from *Fusarium verticillioides* was thermostable and acid stable with broad substrate specificity and high specific activity for animal nutrition purposes. Further according to Vats and Banerjee (2006) that the highly thermostable enzyme and acid stable with wide specificity and high specific activity is an enzyme that potentially to be a co-enzyme and applied in poultry feed. Those it need to biological testing on the feed that phytase enzyme supplemented by *Fusarium verticillioides*.

MATERIALS AND METHODS

Materials used in this study were: (1) 24 male broiler chickens of 4 weeks old. (2) 24 units cage sized 40 x 30 x 30 cm, equipped with incandescent lamps and

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Table 1: Ration composition and feed nutrient (%) and metabolism energy (Kcal/kg) of the treatment ration

No.	Feed ingredient	Treatment				
		R1	R2	R3	R4	R5
1.	Yellow corn	44	44	44	44	44
2.	Rice brain	25	25	25	25	25
3.	Soybean meal	12	12	12	12	12
4.	Fish meal	16.5	16.5	16.5	16.5	16.5
5.	Coconut oil	2.5	2.5	2.5	2.5	2.5
	Total	100	100	100	100	100
6.	Phytase (U/kg)	0	250	500	750	1000
	Nutrient (%)					
	Crude protein	20.03	20.03	20.03	20.03	20.03
	Crude fat	5.59	5.59	5.59	5.59	5.59
	Crude fiber	5.81	5.81	5.81	5.81	5.81
	Calcium	0.98	0.98	0.98	0.98	0.98
	Total phosphor	0.53	0.53	0.53	0.53	0.53
	Available phosphor	0.31	0.31	0.31	0.31	0.31
	Methionine	0.42	0.42	0.42	0.42	0.42
	Lysin	1.26	1.26	1.26	1.26	1.26
	Tryptophan	0.27	0.27	0.27	0.27	0.27
	Metabolism energy (Kcal/kg)	2882.3	2882.3	2882.3	2882.3	2882.3

drinking places. Each experiment unit was occupied by one chicken.

Feed ingredients used to formulate rations for treatment was consists of yellow corn, rice bran, soybean meal, fish meal and coconut oil and phytase. Ration composition and food nutrient and metabolism energy of the treatment ration can be seen in Table 1.

This study used Completely Randomized Design (CRD) with five ration treatments with 4 replications. The treatment were phytase supplementation with dose of: R1: 0 U/kg, R2: 250 U/kg, R3: 500 U/kg, R4: 750 U/kg and R5: 1000 U/kg of ration. Variable measured were retention of phosphor, calcium and nitrogen. The differences between treatments were tested with Duncan's multiple range test (Steel and Torrie, 1991).

RESULTS AND DISCUSSION

The retention of phosphor, calcium and nitrogen are presented in Table 2. Statistical analysis showed that the phytase of *Fusarium verticillioides* supplementation was influenced significantly ($P < 0.01$) on the retention of phosphor, calcium and nitrogen in each treatment during the study.

Retention of phosphorous: DMRT test (Table 1) showed that the retention of phosphor in treatment R1 was highly significant ($P < 0.01$) lower than treatment R2, R3, R4 and R5. But treatment R4 and R5 provides different effects were not significant ($P > 0.05$). It indicated that the higher phytase enzyme addition will be followed by the higher of phosphor retention.

The increasing addition of phytase enzyme on treatment R2, R3 and R4 cause the increase of phosphor retention. This is caused by the supplementation of phytase into the rations which phosphor-deficiency can be improve the utilization of phosphor hydrolysis,

because the phytase will degradation the phosphor which bound in phytic acid. So the phosphor that initially was bound will be utilized. Similar to the research of Rezaei *et al.* (2007) that the supplementation of phytase in poultry rations will increase the availability of phosphor and calcium in broiler. Akyurek *et al.* (2005) stated that phytase supplementation improve the retention of phosphor and calcium in broiler chickens significantly. Furthermore Farrell and Martin (1998) stated the addition of phytase to duck rations containing rice bran has increased significantly of dry matter, nitrogen and phosphor.

Treatments R4 and R5 was not significant. It was indicated that phytase supplementation by 1000 U/kg on treatment R5 did not change the retention of phosphor, because on treatment R4 phytase supplemented up to 750 U/kg of substrate was the optimum substrate hydrolysis by the enzyme and all sides had been occupied by enzyme. So that it became saturated causing phosphor bound with phytic acid could not be hydrolyzed because enzyme could not able to break the phytic acid ties. In the opinion of Zyla *et al.* (2000) that supplementation of 750 U/kg in the ration that wheat was the main ingredients can be increase of weight gain, feed consumption and retention of phosphor and calcium in broiler. Pourreza and Classen (2001) stated that the addition of 500 phytase U/kg can increase the digestibility of protein, phosphor and calcium significantly but the supplementation of phytase up to 1000 U/kg have no effect.

Retention of phosphor on treatment R1 was lower than all treatments because R1 was control, there were not supplemented of phytase. So phosphor which bound to phytic acid could not be hydrolyzed and absorbed by poultry. In addition, the absence of phytase produced in the digestive tract of monogastric livestock, especially

Table 2: Retention of phosphorus (%), calcium (%) and nitrogen (%) of treatment rations

Treatment rations	Retention of phosphor	Retention of calcium	Retention of nitrogen
R1	41.53 ^a	56.69 ^a	55.72 ^a
R2	49.95 ^b	58.53 ^b	64.99 ^b
R3	56.85 ^c	67.98 ^c	66.65 ^c
R4	71.38 ^d	75.65 ^d	67.61 ^d
R5	71.40 ^d	76.98 ^d	68.44 ^d
Mean	58.22	67.17	64.68
SE	0.65	0.46	0.52

Note : Means on the same row with different superscripts are significantly (P<0.01) different.

SE : Standard error.

poultry. According to Mallin (2000), that monogastric livestock, especially poultry do not produce the phytase enzyme in the digestive tract, so phosphor will be excreted with excreta. The results also indicate that the amount of phosphor excreta in treatment R1 was the highest.

Calcium retention: Statistical analysis (Table 2) showed that the retention of calcium in the treatment R1 decreased significant (P<0.01) than the treatment of R2, R3, R4 and R5. The treatment of R4 and R5 were not significant (P>0.05).

Calcium retention of treatments R1 was lower than all treatments because treatment R1 was not supplemented by phytase, so that calcium bound with phytic acid could not be hydrolyzed and absorbed by poultry. According to Pallauf and Rimbach (1996) phytic acid can bind to essential minerals such as Ca, Mg, Fe and Zn. Phytic acid can also bind to amino acids and proteins and inhibit digestion by digestive enzymes, so the enzyme phytase can not be utilized by the body and will be wasted with excreta. This is caused on treatment R1 did not supplemented with phytase, so calcium was bound with phytate could not be hydrolyzed which results in calcium retention of broiler was low. Phytic acid would interfere with mineral absorption valency 2 to Cu, Zn, Mg and Ca, hence, careful measures should be taken for usage in poultry liver (Cullison, 1978).

The increasing addition of phytase enzyme on treatment R2, R3, R4 has increased the calcium retention. This is caused that phytase supplemented into ration with phosphor-deficiency will be improve the utilization of calcium, where phytase will be break down phytase calcium bound to phytic acid that initially bound calcium will be utilized. Similar to the research of Rezaei *et al.* (2007) which states, that supplementation of phytase in poultry rations will increase the availability of mineral phosphor and calcium for broilers. As stated by Akyurek *et al.* (2005) that phytase supplementation can improve significantly the retention of Phosphor and calcium in broiler chickens.

Phytase supplementation of 750 U/kg (R4) and 1000 U/kg (R5) were not increased calcium retention because supplementation of phytase up to 750 U/kg was optimum. In this case all the substat side had occupied

by enzyme, hence, it became saturated. If the addition of phytase increased further to 1000 U/kg the result was not better. Similar results were also obtained by Zyla *et al.* (2000) suggest that supplementation of 750 U phytase/kg in the ration where the wheat is the main ingredients can be increase weight gain, consumption of ration and retention of phosphor and calcium in broiler. Pourreza and Classen (2001) stated that the addition of 500 U phytase/kg can increase the digestibility of protein, phosphor and calcium significantly but the supplementation of 1000 U phytase/kg had no effect further because equaling with phytase supplementation 750 U phytase/kg.

Nitrogen retention: DMRT test (Table 2) showed that the retention of nitrogen in the ration R1 was lower significant (P<0.01) than the treatment of R2, R3, R4 and R5. But the treatment of R4 and R5 were not significant (P>0.05). The increasing phytase supplementation also increased the nitrogen retention but phytase supplementation on ration R5 could not improve nitrogen retention as much as treatment R4.

Nitrogen retention in treatment R1 was lower than other treatments, it caused by treatment R1 without phytase supplemented, so the nitrogen bound to phytic acid could not be hydrolyzed and absorbed by poultry, besides the absence of phytase produced in the digestive tract. According to Mallin (2000) that monogastric livestock, especially poultry do not produce the enzyme phytase in the digestive tract, then the nitrogen was excreted along with excreta. When the nitrogen retention was low the more excreta excreted because many were absorbed by the body, resulting in low value of nitrogen retention and the more nitrogen retention, the less excreta excreted, resulting in high retention value of nitrogen. Anggorodi (1984) states the poor quality of protein in the ration will lead to lower nitrogen retention value or only a small number of proteins that can be used for growth and maintenance.

The increasing addition of phytase enzyme on treatment R2, R3, R4 was followed by increasing nitrogen retention. This is caused that supplementation of phytase into ration with phosphor-deficiency can increase the utilization of nitrogen and energy, because phytase will degradation and breakdown the nitrogen

that bound to phytic acid. So initially nitrogen was bound will be utilized. Phytase enzymes were able to hydrolyzed phytic acid, so that the enzyme phytase can improve the protein or amino acids and energy, as well as calcium and phosphor in feed ingredients (Shelton *et al.*, 2004). In accordance with Selle and Ravindran (2007), phytase supplementation not only improve the digestibility of P, Ca, Mg and Zn but also can directly increase the utilization of nitrogen and energy. Further opinion of Lim *et al.* (2002) stated, that the supplementation of phytase enzyme into the ration can improve significantly digestibility of dry matter, crude fat, P, Zn, Mg and Cu as well as to increase the retention of nitrogen, minerals, Ca, P, Mg and Zn. Furthermore Farrell and Martin (1998a) states the increase phytase supplementation on duck ration contain rice bran has influenced significantly in dry matter, nitrogen and phosphor.

Treatments R4 and R5 not significant. It indicated that phytase supplementation of 1000 U/kg on the R5 did not change the retention of phosphor. It caused treatment R4 with phytase supplemented 750 U/kg had been reached the point of optimum and all the side of substrate was occupied hydrolysis by the enzyme, so that it became saturated. Phosphor bound with phytic acid could not be hydrolyzed because enzyme could no longer afford to break the phytic acid ties. In accordance to Pourreza and Classen (2001) stated that the addition of 500 phytase U/kg can increase the digestibility of protein, Phosphor and calcium significantly but the supplementation of 1000 U/kg had no effect. Nitrogen retention in the ration showed the difference between rations nitrogen consumed with the excreted nitrogen through the excreta, so the higher the nitrogen retention the higher the nitrogen retained in the body (Scott *et al.*, 1982).

The difference of treatment A4 and A5 is not significant because phytase supplementation in treatment A4 can match phytase on treatment A5, where enzyme phytase could hydrolyzed phytic acid. Consequently the protein or amino acids and energy was improved as well as calcium and phosphor in feed ingredients (Shelton *et al.*, 2004). Lloyd *et al.* (1978) states that nitrogen retention is one method for assessing protein quality of the ration by measuring the consumption and excreted nitrogen in excreta, to know the amount of nitrogen retained in the body. Retention of nitrogen in the ration showed the difference between nitrogen consumed with the nitrogen rations secreted through the excreta, the higher retention of nitrogen, the higher nitrogen retained in the body (Scott *et al.*, 1982).

Conclusion: The optimal phytase supplementation on phosphor-deficiency broiler ration was 750 U/kg ration, this can be seen from the retention of phosphor 71.38%, calcium 75.65% and nitrogen 67.61%.

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