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# PRELIMINARY STUDY ON ALLELOPATHIC EFFECTS OF SEVERAL WEED SPECIES ON THE GROWTH OF SOYBEAN (Glycine max (L.) Merr.): THE EFFECT OF OSMOTIC POTENTIAL ON GERMINATION

(Kajian awal allelopati beberapa jenis gulma terhadap pertumbuhan kedelai (Glycine max (L.) Merr.): Pengaruh tekanan osmotik terhadap perkecambahan)

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#### **ABSTRAK**

Percobaan telah dilaksanakan untuk mempelajari pengaruh potensial osmotik larutan ekstrak berbagai jenis gulma terhadap perkecambahan benih kedelai varietas Melrose. Perlakuan adalah berbagai potensial osmotik larutan D-mannitol (0.112, 0.126, 0.131, 0.141, 0.194 and 0.201 Os/kg air) denngan 10 ulangan dalam Rancangan Acak Lengkap (RAL). Konsentrasi larutan D-mannitol tersebut setara dengan konsentrasi larutan ekstrak gulma teki (Cyperus rotundus), bayaman (Amaranthus powellii) dan paspalum (Paspalum dilatatum) yang diekstarksi dengan cara perendaman dan penggilingan. Benih kedelai yang telah disterilisasi-permukaan dikecambahkan pada Petri dish pada suhu 25°C yang diletakkan dalam ruang perkecambahan tanpa cahaya. Jumlah benih berkecambah (dengan panjang radikula ≯ mm) diamati setiap 12 jam selama 84 jam. Pada akhir percobaan dilakukan pengukuran panjang radikula. Hasil percobaan menunjukkan bahwa perbedaan potensial osmotik larutan ekstrak gulma yang berbeda tidak berpengaruh terhadap perkecambahan benih kedelai.

Key words: allelopathy, osmotic potential, soybean, germination

#### INTRODUCTION

bout 10% loss of agricultural production could be attributed to the competitive effects of weeds, in spite of intensive weed control in most agricultural systems (Zimdahl, 1980). Soybean yield losses of 50 to 90% are common for soybean grown in natural weed populations (Coble, 1981), and this yield loss has become serious due to the slow growth of the crop at early stages (Andrade, 1995), highlighting the significance of weed-free competition in soybean. However, knowledge concerning the allelopathic interaction between soybean and some major weeds such as Powell's amaranth (Amaranthus powellii), nutgrass (Cyperus rotundus), and paspalum (Paspalum dilatatum) is also lacking.

Allelopathy refers to any process involving secondary metabolites produced by plants, microorganisms, viruses and fungi that influence the growth and development of

agricultural and biological systems (excluding (Narwal, 1999). Studies demonstrated that Amaranthus powellii, Cyperus rotundus and Paspalum dilatatum interfere with the growth of different soybean cultivars. These weed species had both competitive and allelopathic effects under in vitro and in vivo experimental conditions (Chaniago et al., 2001; 2002; 2003a; 2003b). The inhibition in soybean growth as indicated by total biomass decline might be one of the complex physiological responses of soybean towards weed interference.

Allelochemicals can interfere with germination and radicle elongation through different mechanisms. For example, phenolic acids reduced soybean germination through respiratory enzyme inhibition (Abrahim, 2003), mitochondrial function aberration (Rasmussen, 1992), and plant-water relationships interference (Barkosky, 2003), emphasising that germination inhibition may be a secondary effects of the

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allelochemicals following physiological changes.

To assess the allelopathic effect of weed exudate or leachate on crop species, it is essential that extraction methods mimic the real situation of plant interference. Different extraction methods have demonstrated inconsistent results (either stimulation or inhibition of the target species) (del Moral, 1971; Lovett, 1982; Koch, 1992) causing contention among researchers (Qasem, 1989). For example L-tryptophan, the principal allelopathic compound found in the leachate of mesquite (Prosopis juliflora) foliage, inhibits barnyard grass root growth. However, the amount of Ltryptophan extracted from mesquite foliage using the soaking method decreased significantly with time, whereas the amount of L-tryptophan obtained using a leachate-drip method increased over time (Nakano, 2003).

The work reported here examined the effect osmotic potential and whether different osmotic potential of three weed species (Amaranthus powellii, Cyperus rotundus and Paspalum dilatatum) extracts resulted from

different extraction methods contribute to allelopathy. It is anticipated that this experiment would provide a better understanding of the allelopathic mechanisms and a possible mode of action of three weeds on soybean.

#### **MATERIALS AND METHODS**

Treatments were different osmolality of D-Mannitol (0.112, 0.126, 0.131, 0.141, 0.194 and 0.201 Os/kg water, and purified deionised water as the control treatment), and were replicated 10 times. These ranges of osmolality (Table 1) represent weed extract osmolality of different extraction methods. D-mannitol was used in this study because it has been previously demonstrated to act as an inert osmotic medium for such studies (Wardle, 1992). The pH of each solution and water as the control was adjusted to  $7.5 \pm 0.02$  to meet the average pH of the weed extracts, thus eliminating the effect of different pH on soybean germination.

Table 1. Concentrations and osmolalities of weed extracts at different extraction methods.

Weeds	Extraction methods	Concentrations <sup>1</sup>	Osmolalities <sup>2</sup> (Os/kg water)
Water	N/A	0.0%	0.000
Amaranth	Soaking	3.4%	0.194
	Grinding	3.74%	0.201
Nutgrass	Soaking	2.25%	0.126
	Grinding	2.5%	0.141
Paspalum	Soaking	2.0%	0.112
	Grinding	2.32%	0.131

<sup>1:</sup> concentrations of each extract solutions obtained from the standard curve of standard D-mannitol solutions.

Surface-sterilised soybean seeds were placed in Petri dishes, and supplemented with 10 mL of the D-mannitol solution according to treatments or sterile deionised water. The experiment was conducted in a thermostatically controlled growth cabinet at 25°C in total darkness. The number of germinating seeds (with radicles at least 5 mm long) was recorded every 12 hours for 84 hours, until no further seeds germinated. The radicle length was measured at the end of the experiment. The germination indices were calculated following the formulae of Chiapusio *et al.* (1997).

#### RESULTS AND DISCUSSION

A general trend was observed in which total germination index was not affected by an increase in the osmolality ( $P_{\text{slope}}$ =0.5377,

Figure). The lack of relationship between osmolality level and total germination was also supported by the results of orthogonal contrast between the treatments (P≥0.1491).

Soybean radicle length and radicle dry weight were not affected by different osmotic potentials ( $P_{slope}$ =0.9206 and  $P_{slope}$ =0.8635,

<sup>2:</sup> osmolality values obtained by calculating from known concentrations of D-mannitol.

respectively). In addition, the reduction in radicle length and dry matter accumulation following an increase in osmolality were not significant ( $P_{slope}$ =0.2672 and  $P_{slope}$ =0.2188, respectively; data not shown).

Results from this experiment demonstrated that weed extract osmolality did not affect the germination of soybean. Therefore, adjusting the osmotic potential of weed extracts was not considered necessary in further experiments.

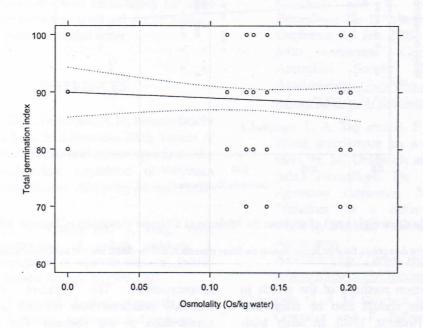


Figure 1. Total germination ( $G_T$ ) index of soybean cv. Melrose at different osmolality of D-mannitol.

The circles indicate the data points, the solid line represents the linear regression, and the dotted lines are the 95% confidence limits.

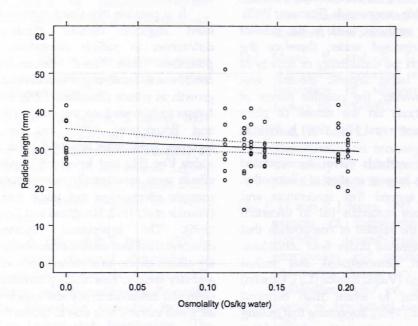


Figure 2. Radicle length (mm) of soybean cv. Melrose at different osmolality of D-mannitol.

The circles indicate the data points, the solid line represents the linear regression, and the dotted lines are the 95% confidence limits.

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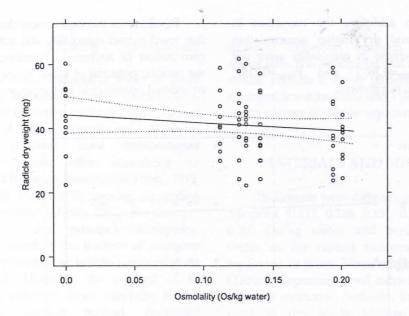


Figure 3. Radicle dry weight (mg) of soybean cv. Melrose at different osmolality of D-mannitol.

The circles indicate the data points, the solid line represents the linear regression, and the dotted lines are the 95% confidence limits.

The extraction methods of the weeds in bioassay studies should also be taken into consideration (Inderjit, 1996). In other trials (data not reported), different extraction methods resulted in similar soybean seed germination responses. However, water extraction neglects the possibility that some organic solvents may enhance the diffusion and activity of soluble compounds (Putnam, 1983). The extraction methods used in the present study used deionised water; therefore the possible effect on the diffusibility or activity of allelochemicals using organic solvents was eliminated. However, the possible effects of extraction methods on the nature of plant compounds (Qasem and Hill, 1989) is difficult to overcome.

Extraction methods have demonstrated different results in past studies of allelopathy. Lovett (1982) argued that maceration and grinding of plant materials led to uncertain results due to the release of compounds that might not be released under field conditions. Another report demonstrated that milled residues of vulpia (Vulpia myuros (L.) J. Gmelin) were more toxic to wheat than chopped residues (An et al., 1997), suggesting that milling plant residues may accelerate the release of free phytotoxins inherent in the material by rupturing cells and increasing the surface area in contact with water. Milling can also provide microorganisms with increased surface area activity which in turn would accelerate the decomposition. The different extraction methods examined here resulted in similar germination in the soybean. The grinding method resulted in a more viscous solution. However, the extract centrifugation separated the supernatant and the aliquot from the extract solution. This may reduce the extract osmolality.

It is possible that weed extracts may also exert negative osmotic effects causing differences in radicle elongation. Osmotic potentials have been demonstrated contribute to reduced germination and seedling growth in lettuce (Bradford, 1990), radish and Lolium multiflorum Lam. var. italicum (Haugland and Brandsaeter, 1996), and the tropical legumes Stylosanthes hamata (L.) Taub and S. scabra Vog (Hu and Jones, 1997). Allelopathic effects were substantially overestimated when osmotic effects were not taken into account (Wardle et al., 1992; Haugland and Brandsaeter, experiment reported The demonstrated that different osmolality had non significant effects on soybean radicle elongation and dry weight. This finding indicates that the observed difference in soybean radicle length in all weed extracts was due to factors other than the extraction methods and the extract osmolality. Reduced radicle growth may therefore have resulted from the allelochemicals derived from the weed extracts.

#### CONCLUSION

The osmotic potentials of the weed extracts did not affect soybean germination. Therefore, adjusting the osmotic potentials of the weed extracts was considered unnecessary for other trials when extraction is carried out by soaking the weed parts in distilled water.

#### REFERENCES

- Abrahim, D., L. Takahashi, A. M. Kelmer-Bracht and E. L. Ishii-Iwamoto, 2003, 'Effects of phenolic acids and monoterpenes on the mitochondrial respiration of soybean hypocotyl axes', *Allelopathy Journal*, 11: 21-30.
- An, M., J. E. Pratley and T. Haig, 1997, 'Phytotoxicity of vulpia residues: I. Investigation of aqueous extracts', *Journal of Chemical Ecology*, 23: 1979-1995.
- Andrade, F. H., 1995, 'Analysis of growth and yield of maize, sunflower and soybean grown at Balcarce, Argentina', Field Crops Research, 41: 1-12.
- Barkosky, R. R. and F. A. Einhellig, 2003, 'Allelopathic interference of plant-water relationships by parahydroxybenzoic acid', *Botanical Bulletin of Academia Sinica*, 44: 53-58.
- Bradford, K. J., 1990, 'A water relations analysis of seed germination rates', *Plant Physiology*, 94: 840-849.
- Chaniago, I., A. Taji and R. S. Jessop, 2001, Interactions of soybean varieties and weed species: Effects of weed density. In Proceedings: The 4th International Legume Conference, 2-6 July 2001, "Legumes Down Under", Australian National University, Canberra.
- Chaniago, I., A. Taji and R. S. Jessop, 2002, In vitro study of weed alelopathy on soybean (Glycine max). In. A. Taji and R. Williams, (eds.) Proceedings: The 7th meeting of the International Association for Plant Tissue Culture and Biotechnology (The Australia Region), 20-23 Jan 2002, "The importance of lant tissue culture and biotechnology in plant sciences", University of New England, Armidale, University of New England Publications Unit, Australia. pp.: 255-262.

- Chaniago, I., A. Taji and R. S. Jessop, 2003a, Assessment of possible alelopathic interactions between soybean (Glycine max) and Amarantus powellii and Cyperus rotundus using in vitro system. In. M. Unkovich and G. O'Leary, (eds.) Proceedings: The 11th Australian Agronomy Conference, 2-6 Feb. 2003, "Solutions for a better environment", Geelong, Victoria, Australian Society of Agronomy, Available: http://www.regional.org.au/au/asa/2003/p/3/chaniago.htm.
- Chaniago, I., A. Taji and R. S. Jessop, 2003b, Weed interference in soybean (Glycine max). In. M. Unkovich and G. O'Leary, (eds.) Proceedings: The 11th Australian Agronomy Conference, 2-6 Feb. 2003, "Solutions for a better environment", Geelong, Victoria, Australian Society of Agronomy, Available: http://www.regional.org.au/au/asa/2003/c/18/chaniago.htm.
- Chiapusio, G., A. M. Sanchez, M. J. Reigosa, L. Gonzalez and F. Pellissier, 1997, 'Do germination indices adequately reflect allelochemical effects on the germination process?' *Journal of Chemical Ecology*, 23: 2445-2453.
- Coble, H. D., F. M. Williams and R. L. Ritter, 1981, 'Common ragweed interference in soybean', *Weed Science*, **29**: 339-342.
- Del Moral, R. and R. G. Cates, 1971, 'Allelopathic potential of the dominant vegetation of western Washington', *Ecology*, **52**: 1030-1037.
- Haugland, E. and L. O. Brandsaeter, 1996, 'Experiments on bioassay sensitivity in the study of allelopathy', *Journal of Chemical Ecology*, 22: 1845-1859.
- Hu, F. D. and R. J. Jones, 1997, 'Effects of plant extracts of *Bothriochloa pertusa* and *Urochloa mosambicensis* on seed germination and seedling growth of *Stylosanthes hamata* cv. Verano and *Stylosanthes scabra* cv. Seca', *Australian Journal of Agricultural Research*, 48: 1257-1264.
- Inderjit, 1996, 'Plant phenolics in allelopathy', The Botanical Review, 62: 186-202.
- Koch, H. J., A. Matthiessen and K. Baeumer, 1992, 'Agronomical risks of straw mulch covers in reduced soil tillage systems .1.

- Influence of chopping intensity of wheat straw on the liberation of phytotoxins', *Journal of Agronomy and Crop Science-Zeitschrift Fur Acker Und Pflanzenbau*, **169**: 184-192.
- Lovett, J. V., 1982, 'The effect of alelochemicals on crop growth and development', In Chemical Manipulation of Crop Growth and Development, (ed.) J. S. Mclaren. Butterworth, London. pp.: 93-110.
- Nakano, H., E. Nakajima, Y. Fujii, K. Yamada, H. Shigemori and K. Hasegawa, 2003, 'Leaching of the allelopathic substance, L-tryptophan from the foliage of mesquite (*Prosopis juliflora* (Sw.) DC.) plants by water spraying', *Plant Growth Regulation*, 40: 49-52.
- Narwal, S. S., 1999, 'Allelopathy in weed management', In *Allelopathy Update, volume 2, Basic and Applied Aspects*, (ed.) S. S. Narwal. Science Publishers, Inc., Enfield, NH, USA. pp.: 203-254.

- Putnam, A. R., 1983, 'Allelochemicals', *Chemical and Engineering News*, 4: 34-45.
- Qasem, J. R. and T. A. Hill, 1989, 'On difficulties with allelopathy methodology', Weed Research, 29: 345-347.
- Rasmussen, J. A., A. M. Hejl, F. A. Einhellig and J. A. Thomas, 1992, 'Sorgoleone from root exudate inhibits mitochondrial functions', *Journal of Chemical Ecology*, **18**: 197-207.
- Wardle, D. A., K. S. Nicholson and M. Ahmed, 1992, 'Comparison of osmotic and allelopathic effects of grass leaf extracts on grass seed germination and radicle elongation', *Plant and Soil*, 140: 315-319.
- Zimdahl, R. L., 1980, Weed-Crop Competition a Review, International Plant Protection Center, Oregon State University, Carvalis.

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