



Test of Randomness of Residuals for the Four-parameter Logistic model used in Obtaining the IC₅₀ Value for *Allivum sativum* Methanolic Extract Against *Aeromonas hydrophila*

Rusnam^{1*}, Gunasekaran, B.² and Sabullah, M.K.³

¹Department of Agricultural Engineering, Faculty of Agricultural Technology, Andalas University, Padang, 25163, Indonesia.

²Faculty of Applied Sciences, UCSI University Kuala Lumpur (South Wing), No.1, Jalan Menara Gading, UCSI Heights 56000 Cheras, Kuala Lumpur, Malaysia.

³Faculty of Science and Natural Resources, Universiti Malaysia Sabah, 88400 Kota Kinabalu, Sabah, Malaysia.

*Corresponding author:

Prof Rusnam

Department of Agricultural Engineering,

Faculty of Agricultural Technology,

Andalas University,

Padang, 25163, Indonesia.

Email: rusnam_ms@yahoo.com

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ABSTRACT

Numerous publications ignore statistical diagnosing of the nonlinear model utilized, and the data might be nonrandom- an essential necessity for all of the parametric statistical evaluation approaches. In cases where the diagnostic tests demonstrate that the residuals reveal a pattern, then a variety of remedies for example nonparametric analysis or shifting to another model should cure the problem. The subject of this study is test for the randomness of the residual for the Four-parameter Logistic model used in obtaining the IC₅₀ Value for *Allivum sativum* methanolic extract against *Aeromonas hydrophila* using the Wald-Wolfowitz runs test. The result shows that the number of runs was 10, the expected number of runs under the assumption of randomness was 5.8, indicating the series of residuals had adequate runs. As the p-value was greater than 0.05, the null hypothesis is not rejected demonstrating no substantial evidence that the residuals were nonrandom, and the residuals represent noise.

INTRODUCTION

Just about the most harmful fish pathogenic agents is *Aeromonas hydrophila*, a known reason behind motile aeromonad septicemia (MAS) in numerous stream fishes as well as being thought to be caused by method of accidental scratches [1-5]. Instances of this ailment especially on fish species are documented in certain great deal places from the United states of America to south East Asia including Indonesia [6]. Species of fish influenced by the bacteria are numerous and include hybrid striped bass, channel cat fish, Goldfish (*Carassius auratus*), Tilapia (*Tilapia nilotica*), Snakehead fish (*Ophiocephalus striatus*), American eel (*Anguilla rostrata*), Carp (*Cyprinus carpio*), Chinook salmon (*Oncorhynchus tshawytscha*) and Rainbow trout (*Oncorhynchus mykiss*) amongst them [7-11]. The bacterium is a Gram-negative rod-shaped and comes from the family Aeromonadaceae. It comes with a great specific polar flagellum that is unbelievably motile, plus it's found in diverse environment such as soil, in sewer, and also in brackish water. The bacterial virulence components consist of its capability to produce a number of tandem-like invasion on the bacterial

system, which includes adhesions, the development of cytotoxins, enzymes like lipases, and the continuing development of a dense biofilm [12-15].

A previous study shows the inhibition of the bacterium *Aeromonas hydrophila* using solvent extracts from *Salvia officinalis* [16]. A nonlinear regression exercise using the four-parameter logistics equation gave the IC₅₀ value of 21.92 mg/mL (95% confidence interval from 20.86 to 23.03). The method of mathematically fitting nonlinear curve using the ordinary least squares method relies heavily on the residuals for the curve to be normally distributed of equal variance (homoscedastic), and does not show autocorrelation [17-19]. Aside from this, an important consideration that has not been highlighted enough is that the residuals must be random. In order for randomness to be met we perform the Wald-Wolfowitz runs test [20] statistical diagnosis tests. The subject of this study is test for the randomness of the residual for the Four-parameter Logistic model used in obtaining the IC₅₀ Value for *Allivum sativum* methanolic extract against *Aeromonas hydrophila* [21].

METHODOLOGY

Acquisition of Data

Data were acquired from **Figure 2** from the works of [21]. Initial outcomes demonstrated that the residuals followed the normally distribution.

Runs test

The runs test [22] was performed to the residuals of the regression in an effort to identify nonrandomness. This might identify an organized deviation of over or under estimation parts of the curve when utilizing a particular model [20]. The runs test compares the series of the residuals which are generally negative and positive. An excellent run is normally signified by an alternation or a balance number of the negative and positive residual values. The number of runs of sign is generally portrayed by means of a percentage of the maximum number feasible. The runs test computes the probability for the existence of way too many or an inadequate number of runs of sign. The existence of too many of a run sign might reveal the existence of negative serial correlation and the existence of too few runs might reveal a clustering of residuals with the exact same sign or the existence of systematic bias.

The test statistic is

H_0 = the sequence was produced randomly
 H_a = the sequence was not produced randomly

$$Z = \frac{R - \bar{R}}{sR} \tag{Eqn. 1}$$

Where Z is the test statistic, \bar{R} is the expected number of runs, R is the observed number of runs and sR is the standard deviation of the runs. The computation of the values of \bar{R} and sR (n_1 is positive while n_2 is negative signs) is as follows;

$$\bar{R} = \frac{2n_1.n_2 + 1}{n_1 + n_2} \tag{Eqn. 2}$$

$$s^2R = \frac{2n_1.n_2(2n_1.n_2 - n_1 - n_2)}{(n_1 + n_2)^2(n_1 + n_2 - 1)} \tag{Eqn. 3}$$

As an example

Test statistic: $Z = 3.0$
 Significance level: $\alpha = 0.05$
 Critical value (upper tail): $Z_{1-\alpha/2} = 1.96$
 Critical region: Reject H_0 if $|Z| > 1.96$

If the test statistic value (Z) is larger than the critical value, then a rejection of the null hypothesis at the 0.05 significance level is made hinting that the sequence was fashioned in a nonrandom manner.

RESULTS

Fitting of a statistical model may be clinically diagnosed precisely using assessments which use residuals. Residuals are the contrast between an expected and observed quantity value of a specific mathematical model. The general rule would be that a poor model will show a bigger difference between the predicted and observed values.

Runs test

From **Table 1**, the number of runs was 10, the expected number of runs under the assumption of randomness was 5.8, indicating the series of residuals had adequate runs. The z-value indicates how many standard errors the observed number of runs is below

the expected number of runs, the corresponding p-value indicate how extreme this z-value is. The interpretation is the same as other o-values statistics. If the p-value is less than 0.05 then the null hypothesis that the residuals are indeed random can be rejected. Since the p-value was greater than 0.05, therefore the null hypothesis is not rejected indicating no convincing evidence of non-randomness of the residuals and they do represent noise.

Table 1. Runs test for randomness.

Runs test	Residual data set
observations	5
below mean	4
above mean	6
no of runs	10
E(R)	5.800
var(R)	2.027
stdev(R)	1.424
Z-value	-0.562
p-value	0.287

The runs test is an important tool to detect nonrandomness in nonlinear regression based on the residuals [22]. The runs test could detect systematic deviation of the curve such as over or under estimation of the sections when using a specific model. The runs test looks at the sequence of the residuals that are usually positive and negative. A good run is usually signified by alternating or a balance number of positive and negative residual values. The number of runs of sign is usually expressed in the form of a percentage of the maximum number possible [20].

In time-series regression models, the runs test is also utilized as a technique to test for the presence of autocorrelation. To be precise, simulation studies using Monte Carlo have shown that the runs test produces distinctly asymmetrical error rates in the two tails suggesting that the use of runs test for autocorrelation detection might not be robust and the Durbin-Watson method would be the method of choice to assess autocorrelation [23]. Previous similar studies based on looking at the randomness of the residuals justify the method use in this study. For instance the use of the Baranyi-Roberts model in fitting an algae growth curve which shows adequacy in the statistics [24]. the Buchanan-three-phase model used in the fitting the growth of *Paracoccus* sp. SKG on acetonitrile [25] and *Moraxella* sp. B on monobromoacetic acid (MBA) [26]. In the arena of biosorption, the residuals for the Sips and Freundlich models utilized in modelling the isotherm of lead (II) uptake by alginate gel bead were found to be adequate based on the runs test [27].

CONCLUSION

The subject of this study is test for the randomness of the residual for the Four-parameter Logistic model used in obtaining the IC₅₀ Value for *Allivum sativum* methanolic extract against *Aeromonas hydrophila* using the Wald-Wolfowitz runs test. The result shows that the number of runs was 10, the expected number of runs under the assumption of randomness was 5.8, indicating the series of residuals had adequate runs. As the p-value was greater than 0.05, the null hypothesis is not rejected demonstrating no substantial evidence that the residuals were nonrandom, and the residuals represent noise.

REFERENCES

1. Ding Z, Zhang Y, Ye J, Du Z, Kong Y. An evaluation of replacing fish meal with fermented soybean meal in the diet of *Macrobrachium nipponense*: Growth, nonspecific immunity, and resistance to *Aeromonas hydrophila*. Fish Shellfish Immunol. 2015;44(1):295–301.

2. Zhou S, Zhang A, Yin H, Chu W. Bacillus sp. QSI-1 modulate quorum sensing signals reduce *Aeromonas hydrophila* level and alter gut microbial community structure in fish. *Front Cell Infect Microbiol.* 2016;6::184
3. Agustina, Prayitno SB, Sabdono A, Saptiani G. Antagonistic activity of kelabau fish (*Osteochilus melanopleurus*) gut bacteria against *Aeromonas hydrophila* and *Pseudomonas* sp. *AACL Bioflux.* 2018;11(6):1859–68.
4. Ahmed HA, Mohamed MEM, Rezk MM, Gharieb RMA, Abdel-Maksoud SA. *Aeromonas hydrophila* in fish and humans; prevalence, virulotyping and antimicrobial resistance. *Slov Vet Res.* 2018;55:113–24.
5. Chen D-D, Li J-H, Yao Y-Y, Zhang Y-A. *Aeromonas hydrophila* suppresses complement pathways via degradation of complement C3 in bony fish by metalloprotease. *Fish Shellfish Immunol.* 2019;94:739–45.
6. Kusumawaty D, Pancoro A, Aryantha INP, Suhandono S. Evaluation of identification techniques for the fish pathogen, *Aeromonas hydrophila*, from Indonesia. *Malays J Microbiol.* 2016;12(3):191–8.
7. Ren S, Guo J, Zeng G, Sun G. Decolorization of triphenylmethane, azo, and anthraquinone dyes by a newly isolated *Aeromonas hydrophila* strain. *Appl Microbiol Biotechnol.* 2006;72(6):1316–21.
8. Stratev D, Vashin I, Daskalov H. Determination of beta-haemolytic activity and minimum inhibitory concentrations of antimicrobial drugs against *Aeromonas hydrophila* strains isolated from fish products. *Bulg J Vet Med.* 2015;18(3):239–47.
9. Tekedar HC, Karsi A, Akgul A, Kalindamar S, Waldbieser GC, Sonstegard T, et al. Complete genome sequence of fish pathogen *Aeromonas hydrophila* AL06-06. *Genome Announc.* 2016;3(2): e00368-15
10. Yang W, Li N, Li M, Zhang D, An G. Complete genome sequence of fish pathogen *Aeromonas hydrophila* JBN2301. *Genome Announc.* 2016;4(1): e01615-15.
11. Rauta PR, Nayak B, Monteiro GA, Mateus M. Design and characterization of plasmids encoding antigenic peptides of Aha1 from *Aeromonas hydrophila* as prospective fish vaccines. *J Biotechnol.* 2017;241:116–26.
12. Ali S, Akhter S, Muhammad A, Khan I, Khan WA, Iqbal MN, et al. Identification, characterization and antibiotic sensitivity of *Aeromonas hydrophila*, a causative agent of epizootic ulcerative syndrome in wild and farmed fish from potohar, Pakistan. *Pak J Zool.* 2016;48(3):899–901.
13. Dias MKR, Sampaio LS, Proietti AA, Yoshioka ETO, Rodrigues DP, Rodriguez AFR, et al. Lethal dose and clinical signs of *Aeromonas hydrophila* in *Arapaima gigas* (Arapaimidae), the giant fish from Amazon. *Vet Microbiol.* 2016;188:12–5.
14. Tkachenko H, Buyun L, Terech-Majewska E, Osadowski Z. In vitro antimicrobial activity of ethanolic extracts obtained from *Ficus* spp. leaves against the fish pathogen *Aeromonas hydrophila*. *Arch Pol Fish.* 2016;24(4):219–30.
15. Velichkova K, Sirakov I, Denev S. In vitro antibacterial effect of *lemna minuta*, *chlorella vulgaris* and *spirulina* sp. Extracts against fish pathogen *Aeromonas hydrophila*. *AACL Bioflux.* 2019;12(3):936–40.
16. Ramena G, Ramena Y, Challa N. Identification and determination of minimum inhibitory concentrations of plant extracts having antimicrobial activity as potential alternative therapeutics to treat *Aeromonas hydrophila* infections. *J Microb Pathog.* 2018 Jan 27;2(1):1–9.
17. Razali NM, Wah YB. Power comparisons of Shapiro–Wilk, Kolmogorov–Smirnov, Lilliefors and Anderson–Darling tests. *J Stat Model Anal.* 2011;2:21–3.
18. Jarque CM, Bera AK. Efficient tests for normality, homoscedasticity and serial independence of regression residuals: Monte Carlo evidence. *Econ Lett.* 1981;7(4):313–8.
19. Snedecor GW, Cochran WG. *Statistical methods.* 7th ed. Ames Iowa: Iowa State University Press; 1980.
20. Motulsky HJ, Ransnas LA. Fitting curves to data using nonlinear regression: a practical and nonmathematical review. *FASEB J Off Publ Fed Am Soc Exp Biol.* 1987;1(5):365–74.
21. Rusnam. Bacterial inhibition activity of methanolic extract from *salvia officinalis*: determination of the IC₅₀ value by nonlinear regression. *Bioremediation Sci Technol Res.* 2018 Jul 31;6(1):23–5.
22. Draper NR, Smith H. *Applied Regression Analysis.* Wiley, New York; 1981.
23. Huitema BE, McKean JW, Zhao J. The runs test for autocorrelated errors: unacceptable properties. *J Educ Behav Stat.* 1996;21(4):390–404.
24. Halmi MIE, Shukor MS, Johari WLW, Shukor MY. Evaluation of several mathematical models for fitting the growth of the algae *Dunaliella tertiolecta*. *Asian J Plant Biol.* 2014;2(1):1–6.
25. Gunasekaran B, Shukor MS, Masdor NA, Shamaan NA, Shukor MY. Test of randomness of residuals for the Buchanan-three-phase model used in the fitting the growth of *Paracoccus* sp. SKG on acetonitrile. *J Environ Bioremediation Toxicol.* 2015;3(1):12–14.
26. Sabullah MK, Shukor MS, Masdor NA, Shamaan NA, Shukor MY. Test of randomness of residuals for the Buchanan-three-phase model used in the fitting the growth of *Moraxella* sp. B on monobromoacetic acid (MBA). *Bull Environ Sci Manag.* 2015;3(1):13–15.
27. Cataldo S, Gianguzza A, Merli M, Muratore N, Piazzese D, Turco Liveri ML. Experimental and robust modeling approach for lead(II) uptake by alginate gel beads: Influence of the ionic strength and medium composition. *J Colloid Interface Sci.* 2014 Nov 15;434:77–88.