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2016 Genetic variation of Rana parvaccola (Inger, Stuart and Iskandar, 2009) based on DNA microsatellites in West Sumatra

by Dewi Imelda Roesma

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Genetic variation of *Rana parvaccola* (Inger, Stuart and Iskandar, 2009) based on DNA microsatellites in West Sumatra

Putri Yuliatmy, Dewi Imelda Roesma and Djong Hon Tjong

Abstract

Rana parvaccola is a new species of differentiated *Rana chalconota* complex in Sumatra. We have analyzed the genetic variation of 30 individuals *R. parvaccola* with Microsatellite DNA in West Sumatra. Five primers were used in the amplification process with PCR technique. The highest expected heterozygosity (He = 0.6533) was Padang population and the lowest (He = 0.2501) was Theorem population. There is a very high genetic differences between populations of *R. parvaccola* in West Sumatra (FsT = 0,3452) with inbreeding coeffesient value Fis = -0,4152 and FiT = 0.0849. The value of gene flow between populations was very low (Nm = 2.0613).

Keywords: genetic variation, Microsatellite DNA, Rana parvaccola, gene flow

1. Introduction

Rana parvaccola is an amphibian species that identified as new species from *Rana chalconota* complex in Sumatra. R. chalconota is a species that is widespread in the Sundaland ^[1]. Geographical isolation can lead to changes morphotype between populations at the complex species such as size and color pattern ^[1-3]. *R. rufipes* and *R. parvaccola* are species of R. chalconota complex only found in Sumatra. *R. rufipes* is a large R. chalconota, have red swimming membrane, size of the tympanum is equal to the diameter of the eye in females and larger in males which on the inside a little depressed. *R. parvaccola* is a small R. chalconota, have black swimming membrane, tympanum approximately two-thirds the size of the diameter of the eye in females and larger in males ^[1]. Based on surveys and observations that have been done in West Sumatra, we know that *R. parvaccola* found frequently than the *R. rufipes*. Out of nine locations that have been surveyed we found *R. parvaccola* at six locations, while *R. rufipes* found in three locations. *R. parvaccola* found in habit an area with various heights and regions of West Sumatra.

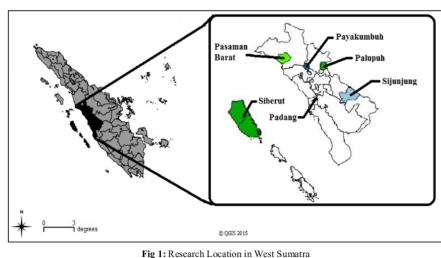
Morphological form and genetic constitution of amphibian were affected by human-caused habitat destructions, altitude landscape and geographical factors ^[4-6]. Amphibia spend their entire life cycle in water and land so they are generally believed to be sensitive to environment perturbations, partly because of their central place in the food chain. Land use changes could be the most critical factor for the population decline and it's has been reported in various parts of the world ^[6, 7]. Microsatellite markers are used in interpopulation diversity and gene flow studies of natural populations and also provide relevant information for identifying conservation units and for investigating the genetic processes that take place in populations such as the incidence of genetic drift and generation of genetic neighborhood ^[8, 9]. According o the topography of West Sumatra we assumed genetic variation in population of *R. parvaccola*. This study aimed to analyze the genetic variation intra- and inter-population *R. parvaccola* in West Sumatra.

2. Materials and Method

Individual of *Rana parvaccola* and D1A samples collected during March-June 2013 at six locations in West Sumatra (Figure 1). Analysis of genetic variation used DNA Microsatellite markers^[2, 10, 11]. We used the QIAGEN kit protocol DNAeasy® Blood and Tissue Kit (QIAGEN, Inc., Valencia, California, USA) for the DNA isolation. The quality of DNA was tested by 1,2% gel electrophoresis added 3 µl Ethium Bromida (EtBr). Amplification of DNA was programmed for 33 cycles with modified temperature and time (denaturation: 950C;120

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minutes, annealing: modification time; 30 minutes, extention: 72ºC;10 minutes) using PROMEGA PCR Core Kit. Nine primers were used (RNH1, RNH2, RNH3, RNH4, RNH6, RNH9, RNH10, RNH12 and RNH13) refers to the Gong ^[12] with modification. Identification DNA amplification used 2% **1** electrophoresis added 5 μl Ethium Bromida (EtBr). Quantification of the results analyzed using GENEPOP ver 4.0 program to determine the genetic variation within and between populations. The parameters used to determine the genetic variation within a population were, allele frequencies, the percentage of polymorphic loci (Pp), the average number of alleles per lokus (Na) and expected heterozygosity (HE). Genetic variation between populations can be seen from the Fstatistic, genetic distance and similarity values dendogram based on Nei's genetic distance matrix (1978) and the value of gene flow (Nm) [13].



1										
Т	able 1:	Geographic	position a	and	altitude	of	reasearch	location	of R.	parvaccola

No	1 Location	District	Altitude (mdpl)	Geographic Position
1.	Cagar Alam 1 mbah Harau-Payakumbuh	50 Kota	500-600	0° 04'LS dan 100°39' BT
2.	1 Pasaman Barat	Pasaman Barat	456-763	0º 11' LS dan 100º 04' BT
3.	Hutan Pendiditan Penelitian Biologi (HPPB) Unand Padang	Padang	260-300	0º21' LS dan 100º 46' BT
5.	1Cagar Alam Pangean I-Sijunjung	Sawahlunto- Sijunjung	300-600	0º 90'LS dan 101º 50' BT
6.	Cagar Alam Batang Palupuah-Agam	Agam	700-1000	0° 14' LS dan 100°21' BT

3. Results and Discussion

Based on the results of selection of the nine primers, we found that only five primer produces the amplicon that is the primer RNH3, RNH9, RNH10, RNH12 and RNH13. The number 11 alleles at each locus ranged from 4-13 alleles with total of as much as 41 alleles and an average of 8.2 alleles per primer. All alleles obtained a polymorphic alleles. A locus is usually considered to be polymorphic if the ferquency of the most common allele is less than either 95% or 99% [14, 15]. The value of R. parvaccola expected heterozygosity intra-population in West Sumatra ranged from 0.4044 to 0.6533 with the percentage 1 f polymorphic loci (Pp) ranged from 80-100 (Table 2). Based on that value we know that population of Padang, Sijunjung, and Pasaman Barat have a high genetic variation while population of Payakumbuh, Palupuh and Siberut have a lower. The population that has the highest heterozygosity values is population of Padang with a value of heterozygosity (He) of 0.6533 and Shannon Index value (I) \pm 0.0911 and a population that has the lowest heterozygosity

values is Siberut population (H = 0.4044; I = 0.2501). High heterozygosity in the population due to large population sizes so that mating occurs randomly. In lin 15, 16 a large population size will suppress the occurrence of inbreeding and increase mating randomly so that genetic variation and heterozygosity in the population will increase.

The lowest heterozygosity found in Siberut population. Siberut is an island separated from the 2ainland of Sumatra by the Indian Ocean. According to [17] reduced genetic diversity on oceanic islands is expected because island populations are typically founded by a few migratory individuals, resulting in extreme genetic bottlenecks. These conditions causes increase of inbreeding on the Siberut population. Low levels of genetic variation can be expected based on small population sizes in the islands. Island acts as a barrier to dispersal and migration of amphibians. Therefore, the effects of drift will be intensified when founding populations are small, when islands are small in area and when islands are far from the mainland source population [17, 18].

Table 2: Intra-population genetic variation of R. parvaccola based on DNA microsatellite

Population	Na	Ne	I	He	Pp (%)	N
Padang	3,0000 ±1,0000	2,5028 ±0,4761	0,9174 ±0,2657	0,6533 ±0,0911	100	5
Payakumbuh	2,0000 ±0,7071	1,8204 ±0,6160	0,5833 ±0,3776	0,4311±0,2688	80	4
Palupuh	3,4000 ±1,9494	2,7765 ±1,4796	0,9606 ±0,6435	0,5822 ±0,3462	80	4
Sijunjung	3,2000 ±1,6432	2,5393 ±0,7551	0,9714 ±0,3816	0,6444 ±0,1299	100	5
Siberut	2,2000 ±1,0954	1,7151 ±0,5234	0,5787 ±0,3923	0,4044 ±0,2501	80	4
Pasaman Barat	2,6000 ±0,8944	2,2819 ±0,3894	0,8555 ±0,2276	0,6133 ±0,0795	100	5

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Description : Na : the average number of alleles perlokus; Ne : the average of efective alleles; I : the average of Indeks Shanon (Lewontin, 1972); He : heterozigosity (genetic diversity Nei); PP: the percentage of polymorphic loci; N: Number of polymorphic loci Inter-population genetic structure of R. parvaccola based on microsatellite DNA can be described by the total inbreeding value (FIT: 0.0849), inbreeding intra-population (FIS: -0.4152), coefficient of genetic differentiation (FST: 0.3534) and value gene flow (Nm: 0.4575). FIS positive indicates the decrease of heterozygosity intra-population and negative FIS indicate increased heterozygosity intra-population. So, we know that the heterozygosity intra-population is higher than heterozygosity inter-populations. F_{II} value indicated that the mating of R. parvaccola among population is low. The value of generation differentiation between populations (F_{ST}) can also be used to determine the genetic variation between populations. Value of the $F_{ST} > 0.25$ indicates a very high genetic differentiation ^[15]. Genetic differentiation inter-population of R. parvaccola in

West Sumatra determined by F_{ST} value, known that the genetic variation interpopulation lower than intrapopulation.

Levels of migration or gene flow is high if value of Nm>1 [15, ^{19]}. Our results also show that the gene flow (Nm) between populations is low (0.4575; Nm \leq 0.5). Gene flow is caused by the movement or dispersal of whole organisms or genomes from one population to another. After entering a new population, immigrant genomes may become incorporated due to sexual reproduction or hybridization, and will be gradually broken up by recombination, so can reduce the genetic differentiation between populations and increases genetic variation in the population [15, 20]. From the value of gene flow in populations of R. parvaccola in West Sumatra we predicted that the migration of individuals between populations is low. This condition make the population of R. parvaccola in West Sumatra devided into subpopulation. The dendogram of R. parvaccola population in West Sumatra showed in Figure 2 and the genetic distance Nei 1978 showed in Table 3.

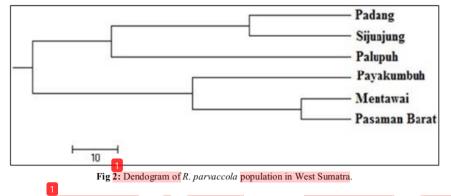


Table 3: Nei genetic distance matrix 1978 in six populations of R. parvaccola based on microsatellite DNA markers.

No.	Population	1	2	3	4	5	6
1	Padang	-					
2	Payakumbuh	1,4314	-				
3	Palupuh	0,9457	0,9251	-			
4	Sijunjung	0,5253	2,2031	1,2988	-		
5	Siberut	1.7838	0,7389	1,2864	1,3061	-	
6	Pasaman Barat	1,4986	0,7393	1,5299	1,1599	0,2638	-

The grouping of *R. parvaccola* by UPGMA cluster analysis and the value of genetic distance between populations (Table 3) more than 0.5 (D> 0.5) indicate that the genetic structure of populations *R. parvaccola* in West Sumatra have been separated ^[2, 15, 21]. We preddicted that microhabitat factors such as habitat structure damage, antropogenic and type of vegetation change causeing genetic differentiation.in *R. parvaccola* We suggest to study about microhabitat and ecological factors of *R. parvaccola*.

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5. References

 Inger RF, Stuart BL, Iskandar DT. Systematics of a Widespread Southeast Asian frog, *Rana chalconota* (Amphibia: Anura: Ranidae). Zoological Journal of the Linnean Society. 2009; 155(1):123-147.

- Wang IJ, Kyle S. Genetic Structure is Correlated With Phenotypic Divergence Rather Than Geographic Isolation In The Highly Polymorphic Strawberry Poison-Dart Frog. Molecular Ecology. 2010; 19(3):447-458.
- Wang Y, Jian Z, Jianhuan Y, Zhixin Z, Guoling C, Yang L. Morphology Molecular Genetics and Bioacoustic Support Two New Sympatric Xenophrys Toads (Amphibia: Anura: Megophrydae) inSoutheast China. Plos one. 2014; 9(4):1-15
- Johansson M, Primmer CR, Sahlsten J, Merila J. The Influence of Landscape Structure on Occurance, Abundance and Genetic Diversity of The Common Frog *Rana Temproraria*. Global Change Biology. 2005; 11(10):1664-1679.
- Schauble C. Variation in The Body Size and Sexual Dimorphism Across Geographical and Environmental Space in The Frogs *Limnodynastes tasmaniensis* and *L. peronei*. Biological Journal of the Linnean Society. 2004; 82:39-54
- Scribner KT, Arntzen JW, Burke T, Cruddace N, Oldham RS. Environmental correlates of Toad abundance and

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population genetic diversity. Biological Conservation. 2001; 98(2):201-210.

- Allentoft ME, dan John O. Global Amphibian Declines, Loss of Genetic Diversity and Fitness: A Review. Diversity. 2010; 2(1):47-71
- Oliveira EJ, Padua JG, Zucchi MI, Venckovsky R, Vieira MLC. Origin, Evolution, and Genom Distribution of Microsatellite. Genetics and Molecular Biology. 2006; 29(2):294-307.
- Heywood VH, Iriondo JM. Plant Conservation: Old problems new perspective. Biological Conservation. 2003; 113:321-335.
- Arens P, Bugter R, Westende WV, Zollinger R, Stronks J, Vos CC, Smulders MJM. Microsatellite variation and poplation structure of a recovering Tree Frog (*Hyla arborea* L) metapopulation. Conservation Genetics. 2006; 7(6):825-835.
- Funk WC, Michael SB, Paul SC, Bryce AM, David SP, Stephen A et al. Population structure of Columbia spotted frogs (*Rana luteiventris*) is strongly affected by the landscape. Molecular Ecolology. 2005; 14(2):483-496.
- Gong J, Hong L, Sheng GF, Qiu HW. Development and characterization of 13 Polymorphic Microsatllite DNA markers for the pond green frog (*Rana nigromaculata*). Journal of Genetic. 2010; 8(1):e7-10.
- Raymond M, Rousset F. GENEPOP (version 4.1.10): Population Genetics Software For Exact Test and Ecumenicism. Heredity. 1995; 86:248-249.
- Finkeldey R. An Introduction to Tropical Forest Genetics. Edition I, Institute of Forest Genetics and Forest Tree Breeding Publishing, Gottingen, 2005, 31-32.
- Allendorf FW, Gordon HL. Conservation And The Genetics Of Populations. Edition I, Blackwell Publishing, USA, 2007, 197-327.
- Phillipsen IC. Population genetics of Ranid frogs: Investigating effective population size and gene flow. Ph.D. Disertation. Oregon State University. USA. 2001.
- Duryea MC, Zamudio KR, Brasileiro CA. Vicariance and Marine Migration in Continental Island Populations of a Frog Endemic to The Atlantic Coastal Forest. Heredity. 2015; 115(3):225-234
- Seppa P, dan Anssi L. Genetic structure of island population of the anurans *Rana temporaria* and *Bufo bufo*. Heredity. 1999; 82(1):309-317.
- Slatkins M. Estimating Levels of Gene Flow in Natural Population. Genetics. 1981; 99(2):323-335.
- Mallet J. Gene Flow. In: Woiwod IP, Reynolds DR, and Thomas CD (Eds.) Insect Movement: Mechanism and Consequences. CAB International, Wallingford, UK, 2001, 337-360.
- Silva DDME, Aparecido DDC, Rogerio PB, Mariana PCT, Jose AFDF. Population structure of *Eupemphix nattereri* (Amphibia, Anura, Leiuperidae) from Central Brazil. Genetics and Molecular Biology. 2008; 30(4):1161-1168.

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