

Turnitin Originality Report

Processed on: 28-Jan-2020 10:09 PM +08

ID: 1247659262

Word Count: 3939

Submitted: 1

JWPR2019 By Jwpr Jwpr

3% match
(publications)

[Mirnawati ., Ade Djulardi, Gita Ciptaan. "Utilization of Fermented Palm Kernel Cake with Sclerotium rolfsii](#)

[in Broiler Ration", International Journal of Poultry Science, 2018](#)

Similarity Index	Similarity by Source	
9%	Internet Sources:	N/A
	Publications:	9%
	Student Papers:	N/A

3% match (publications)

[Mirnawati ., Gita Ciptaan, Ade Djulardi. "The Combined Effects of Fungi Phanerochaete chrysosporium and Neurospora crassa and Fermentation Time to Improve the Quality and Nutrient Content of Palm Oil Sludge", Pakistan Journal of Nutrition, 2019](#)

2% match (publications)

[Dwi Utami Anjarwati, Rizka Hidayati, Dian Kristiantoro, I. D. S. A. P. Peramiarti, Ari Asnani. "The protein content and protease activity of local green fly, Chloroprocta sp., maggot crude extracts", AIP Publishing, 2019](#)

1% match (publications)

[Abiala Abiala Alatise. "chapter 3 The Impact of Palm Kernel Cake in Resolving Herdsmen-Farmers Violent Conflicts in Ogun State, Nigeria", IGI Global, 2019](#)

1% match (publications)

[Ulvi Fitri Hand, Wizna ., Irfan Suliansyah, Yose Rizal, Maria Endo Mahat. "Effects of Heating Method on Lycopene, Dry Matter and Nutrient Content of Tomato \(Lycopersicon esculentum\) Waste as Laying Hen Feed", International Journal of Poultry Science, 2018](#)

JWPR 2019, Scienceline Publication J. World Poult. Res. 9(4): 211-216, December 25, 2019 Journal of World's Poultry Research Research Paper, PII: S2322455X1900027-9 License: CC BY 4.0 DOI: <https://dx.doi.org/10.36380/jwpr.2019.26> **The Effect of Bacillus subtilis Inoculum Doses and Fermentation Time on Enzyme Activity of Fermented Palm Kernel Cake** Mirnawati1*, G. Ciptaan1 and Ferawati1

Animal Nutrition Department, Animal Science Faculty, Andalas University, Padang 25163, Indonesia *Corresponding author's Email: mirnawati@ansci.unand.ac.id; ORCID: 0000-0002-9887-7227

Received: 29 Oct. 2019 Accepted: 06 Dec. 2019 ABSTRACT [Palm kernel cake \(PKC\)](#) was [by-product of palm oil](#) industry and it had potential to be one of the poultry ration ingredient. However, its utilization for poultry was still limited because of the β -mannan in PKC. In order to increase PKC utilization in poultry ration, fermentation process was done to remodeled β mannan by using *Bacillus subtilis*. This research conducted a study on the effect of *Bacillus subtilis* inoculum dose and fermentation time to increase the enzyme activity of FPKC by using CRD with 3×3 factorial and 3 replications. Factor A was 3 doses of inoculum *Bacillus subtilis*: 3%, 5%, and 7%. Factor B was fermentation times which contained: (1) 2 days, (2) 4 days, and (3) 6 days. Parameters used were enzyme activity of mannanase, protease, and cellulase in FPKC. Significant interaction was seen between inoculum doses of *Bacillus subtilis* and fermentation time. There was also a significant interaction on each of the inoculums dose of *Bacillus subtilis* and fermentation time on all of the enzyme activity. This study concluded FPKC with *Bacillus subtilis* of 7% inoculums doses and 6 days fermentation time indicate the best result as seen from 24.27 U/ml of mannanase activity, 10.27 U/ml of protease activity, 17.13 U/ml of cellulase activity of fermented PKC. Key words: *Bacillus subtilis*, Enzyme activity, Fermentation time, Inoculum doses, Palm Kernel Cake

INTRODUCTION The availability of feed ingredients is still a common problem experienced by farmers, because of its insufficient availability. Various ways have been tried in order to availability become sufficient. One of the efforts to overcome that problem is by using plantation waste as a cheaper alternative feed ingredient, while not competing with human needs, such as palm kernel cake.

Indonesia as [the largest palm oil producer in the world](#), has produced 45 million tons of palm oil/year (DGP, 2016). In West Sumatra, oil palm plantations have an area of 399,120 hectares, producing 1,145,432 tons palm oil per year (Central Bureau of Statistics, 2015). Each of fresh palm bunches produces 5% of palm kernel and it also produces 45-46% of palm kernel cake or 2.0-2.5% by weight of palm bunches. Nutritional content of palm kernel cake is 17.31% crude protein, 7.14% crude fat, 27.62% crude fiber, [0.27% Ca, 0.94% P and 48.4 ppm Cu](#) (Mirnawati et al., 2018). The [utilization of palm kernel cake in broiler](#) rations is limited to only 10% (Rizal, 2000). Low utilization of [palm kernel cake in poultry](#) rations is [due to its high mannan content](#). As stated by Daud and Jarvis (1993). The 56.4% of crude fiber content [in palm kernel cake is in a form of](#) β -mannan. However, poultry do not have β -mannan hydrolysis enzyme in their bodies. In addition to the high crude fiber content in palm kernel cake, its low protein and amino acid digestibility also caused low utilization of palm kernel cake. As well as, Tafsir (2007) found that the low utilization of palm kernel meal in poultry rations is caused by the high content of crude fiber, low digestibility of proteins and amino acids. Fermentation is one of the methods to improve [the quality of the palm kernel cake. Fermentation](#) is a change in chemical material of feed ingredients because of the enzymes produced by

microorganisms or existed in these feed ingredients (Buckleet al., 1987; Rizal et al., 2013). Fermented palm kernel cake was done with the help of mannanolytic fungus, for it can produce mannanase enzyme that hydrolyzes mannan. Mirnawati et al. (2017) studied palm kernel cake fermentation using mannanolytic fungus such as *Sclerotium rolfsii*, *Eupenicillium javanicum* and *Aspergillus niger*. The *Sclerotium rolfsii* has a higher ability among the three fungus to produce enzyme [To cite this paper: Mirnawati, Ciptaan G and Ferawati \(2019\). The Effect of Bacillus subtilis Inoculum Doses and Fermentation Time on Enzyme Activity of Fermented Palm Kernel Cake. J. World Poult. Res., 9 \(4\): 211-216. http://jwpr.science-line.com](#) 211 compared to *Eupenicillium javanicum* and *Aspergillus niger*. *Sclerotium rolfsii* has mannanase enzyme activity of 24.58 U / ml and cellulose activity of 21.89 U / ml and can improve the [nutritional quality of palm kernel cake, as](#) seen from 23.66% of crude protein, 16.72% of Crude fiber, 0.22% of crude lipid, 0.75% of Calcium, 0.85% of Phosphor, 57.16% of nitrogen retention, and 25511 kcal/kg metabolizable energy of palm kernel cake. Although there has been an increase in the nutritional content [of fermented palm kernel cake with Sclerotium rolfsii](#), its utilization [in broiler](#) rations is only up to 25% (Mirnawati et al., 2018). Fermentation with the help of fungi takes longer time than microbes. According to Fardiaz (1992), microbes as an inoculum requires less time than fungi in the fermentation process, which is about 1-2 days, because the division time is faster. In addition, there are also mannanolytic microbes which are *Bacillus subtilis* WY34 (Jiang et al., 2006). Hooge (2003) added that *Bacillus subtilis* can produce several enzymes such as protease, β - mannanase and several enzymes that are useful in helping digestion so that it is easier to digest. *Bacillus subtilis* is known to be capable of producing cellulase when placed in an environment containing cellulose. *Bacillus subtilis* can produce protease, α -amylase, and renin (Darwis and Sukara, 1990). Many factors including dose of inoculum and length of fermentation need to be considered following the fermentation process. The application of the right inoculums dose will provide opportunities for rapid growth and development of microbes when more doses of inoculum used, the fermentation process occurs faster, thus more substrates changed. Furthermore, the longer fermentation period lasts, the more substances that are remodeled (Fardiaz, 1992 and Mirnawati et al., 2013). Therefore, it is necessary to know the optimal dose and duration [of palm kernel cake](#) fermentation [with Sclerotium rolfsii to](#) produce maximum enzyme activity to increase the nutritional value of palm kernel cake. [MATERIALS AND METHODS The materials used in the study were: palm](#) kernel cake, fine bran, *Bacillus subtilis* InaCC B289, media (NA/ Nutrient Agar), distilled water, buffer solution, and chemicals for analysis of mannanase activity, cellulase and protease. The tools were autoclaved and analytical scales were made in Japan. Incubators, spectrophotometers, and shakers waterbath are made in Jerman. Centrifuges, erlenmeyer, pH meters were made in China. This research was conducted using experimental methods using [a completely randomized design \(CRD\), a factorial pattern](#) consisting of 2 treatment factors, namely factors A and B with three replications (Steel and Torrie, 1991). Factor A is the dose of inoculum, namely:

A1 = 3%, A2 = 5% and A3 = 7%. Factor B is the duration of fermentation, namely: B1 = two days, B2 = four days and B3 = six days. Measured variables were the activity of Mannanase, cellulase and Protease. Fermentation carried out with a ratio of 80:20 palm kernel meal and bran with 60% moisture content. Materials autoclaved for 15 minutes at 121°C, Chilled at room temperature then *Bacillus subtilis* inoculated on ingredients. After inoculation, *Bacillus subtilis* then fermented in the incubator. Enzyme extract 10 g of the fermented sample, then soak with 90 ml of 0.05 M pH buffer phosphate. Leave it in the shaker incubator at 100 rpm for 30 minutes. After that, strain with filter paper and take the filtrate. The filtrate obtained was centrifuged with 10,000 rpm, temperature of -4°C for 15 minutes. Take the supernatant and the enzyme activity will be analyzed. Enzyme activity measurement ? Mannanase and protease by N-Somogyi Nelson Method Mannanase and protease by N-Somogyi Nelson Method One ml of crude enzyme substrate was added to one ml of manan substrate (0.5 g/ml manan plus 10 ml of phosphate buffer). Incubate for 30 minutes at temperature (40 cellulases and 60 mannanases) in the waterbath shaker. Take one ml extract of the enzyme that has been incubated, and then add the Nelson AB solution. Heat in boiling water for 20 minutes, after that cool and add one ml of phosphomolibdat solution and seven ml of distilled water measure with a Uv-Vis spectrophotometer wavelength 575 nm. ? Protease Protease The crude extract of the extracted enzyme determined by its proteolytic activity based on Cupp and Enyard (2008). One [ml of crude enzyme extract was added to the casein substrate 0.65% \(0.65 g casein in 100 ml Pospat buffer 0.05 M pH 7.5\). The reaction mixture was incubated at 37° C for 10 minutes. Termination of the reaction was carried out](#) by adding five ml of 110 ml TCA reagent, and re-incubating [at 37° C for 30 minutes. Two ml of filtrate was separated by centrifugation at 10000 rpm for 10 minutes. Five ml of Na2CO3 and one ml of Folin J. World Poultr. Res., 9\(4\): 211-216, 2019 Ciocalteau reagent were added to the filtrate and incubated at 37° C for 30 minutes. The absorbance of the mixture was measured using a UV-vis spectrophotometer at a wavelength of 660 nm.](#) Data analysis Datas obtained were processed statistically by analysis of variance of [a completely b randomized design \(CRD\) with factorial](#) patterns 3x3 [with](#) tree replicates. Differences between treatments test [by using Duncan multiple range test \(DMRT\) \(Steel and Torrie, 1995\). RESULTS AND DISCUSSION Treatment effect](#) on mannanase activity The mean mannanase activity of fermented PKC with *Bacillus subtilis* was illustrated in table 1. Table 1 showed that there was a tendency of increase in mannanase activity along with the addition of the inoculum dose. Increasing doses of inoculum caused the higher the activity of mannanase on 2 days, 4 days and 6 days of fermentation time. Furthermore, the longer fermentation time increases mannanase activity, at the inoculum dose of 3%, 5% and 7%. The above data could be concluded that the 7% inoculum and 6-days fermentation time (A3B3) provided the highest mannanase activity. The high mannanase activity of A3B3 (6.27 U/ml) was concluded by high doses of inoculum and the longer time of fermentation given to microorganisms allows it to grow and develop rapidly, thus increased mannanase activity. Consistent with Mirnawati

et al. (2017) that reported giving the more doses of inoculum caused faster fermentation process, because of the high doses of inoculum that enables increase of enzyme activity and microbial growth in the substrate. Fardiaz (1992) found that the slow pace of fermentation greatly determined the amount of enzymes produced in the media. The longer the fermentation time occurs, the more substrate will be degraded by enzymes produced by microbes. The low mannanase activity in the A1B1 treatment (15.49 U/ml) was caused by smaller dose of inoculum, which is at 3% and short duration of fermentation (2 days). This caused the slow growth of microbes and low activity of enzyme, however with an increase in the inoculum dose of 7%, the growth of microbes will be better. So, the dose of 7% inoculum is the optimum doses for the growth of *Bacillus subtilis*. In accordance with Darwis et al. (1995) at the beginning of fermentation, enzyme activity was very low and it will increase along with the increasing fermentation time. The enzyme activity follows the growth pattern, starts from the phase of adaptation, exponential, stationary and death phase. Table 1. Mean mannanase activity fermented palm kernel cake with *Bacillus subtilis* Factor A Factor B (Fermentation time) (Inoculum Dose) B1 (2 days) B2 (4 days) B3 (6 days) Mean A1 5.49aAB 5.62aB 5.52aC 5.55 A2 5.60bB 5.66bAB 5.94aB 5.73 A3 5.80bA 5.81bA 6.27aA 5.96 Mean 5.63 5.70 5.91 -- Note: Different uppercase letters in different columns and small letters on the same row showed very significant different (P < 0.05). Mean mannanase activity is in U/ml.

Treatment effect on cellulase activity The mean cellulase activity of fermented PKC with *Bacillus subtilis* is shown in table 2. There was an increase of cellulase activity as seen from table 2, along with the addition of the inoculum doses. Cellulase activity was increased at fermentation times (2 days, 4 days, and 6 days), proving that more length of fermentation will increase cellulase activity. Inoculum doses influenced by cellulase activity, evidenced by an increased in cellulase activity with more inoculum doses of 3% 5% and 7% inoculum doses. From the above data, it can be concluded that the 7% inoculum dose and 6 days fermentation time (A3B3) provides the highest cellulase activity at 16.11 U/ml. The high cellulase activity of A3B3 treatment at 16.11 U/ml caused by the increasing doses of inoculum and the length of fermentation given which allows the rapid growth and development of microorganisms, so the cellulase activity will be increased. In accordance with the opinion of Mirnawati et al. (2013) that higher dose of inoculum provided better environment for microbial growth causing faster fermentation process while also increasing the enzyme In addition, fermentation time is also one of the determinant factor, where longer fermentation time will cause more remodeled substrate that produced by enzyme of microbes (Mirnawati et al. 2012). The low cellulase activity in the A1B2 treatment (5.49 U/ml) caused by low doses of inoculum at 3% and 4 days of fermentation time. At this treatment, microbes tend to growth slowly and the enzyme activity will be lower. However, the increase in the inoculum dose of 7% was causing microbes to grow better, thus 7% inoculum dose is recommended as the optimal dose for the growth of *Bacillus subtilis* which is in accordance with Zulfatus et al. (2008) that enzyme activity was obtained at post exponential (stationary) time after the 4th day of fermentation. At the incubation

period, it was shown that the cellulose enzyme worked optimally in hydrolyzing the substrate, namely cellulose found in palm kernel cake, into glucose. Treatment effect on protease activity The mean protease activity of fermented PKC with *Bacillus subtilis* is shown in table 3. There is a tendency of an increase in protease activity as presented in table 3, along with the supplementation of inoculum dose. The increased doses of inoculum come with the increased protease activity, both at fermentation times (2 days, 4 days and 6 days). Longer time of fermentation also increases the protease activity, at the inoculum dose of 3%, 5% and 7%. It can be concluded from the above data that the 7% inoculum dose and 6 days of fermentation time (A3B3) shows the highest protease activity at 10.27 U/ml. The highest protease activity is shown from A3B3 treatment, it was caused by the increased dose of inoculum and the longer fermentation time, so microorganisms may grow and develop more while also increasing protease activity. [In accordance with the opinion of MUSAALBAKRI ET AL. \(2005\)](#), the number of inoculums has effect on increasing cell concentration gradually because it increases microbial growth. Thus by increasing the inoculum, the microbes will produce more protease enzymes. DADA ET AL. (2009) also found that high doses of inoculum added would increase the metabolic compounds produced, while higher inoculum concentrations were also inefficient when the fermentation process was carried out. The longer the fermentation time, the higher the enzyme activity produced. According to DWIDJOSEPUTRO (2010) that there is an incubation time in producing metabolic compounds of each bacterium according to the growth phase of each bacterium. The low protease activity in the A1B3 treatment at 5.36 U/ml was caused by a small inoculum dose at 3%, resulting in low microbial growth and enzyme activity. The increasing dose of inoculums will provide a better environment for the growth of microbes, thus 7% inoculum is the optimum dose for the growth of *Bacillus subtilis*. This is in accordance with the statement of BELMA ET AL. (2000) that reported incubation time affects the cell growth process which occurs through cell division. That process will increase the living cells and the rate of growth in its culture. During the incubation period, it was shown that protease enzymes performed optimally in hydrolyzing the substrate, namely the protein found in palm kernel cake into amino acids and protein quality increases (MIRNAWATI ET AL., 2019).

Table 2. Mean cellulase activity of fermented palm kernel cake with *Bacillus subtilis*. Factor A (Inoculum Dose) B1 (2 days) 12.34bB 12.33bC 13.15aC [Factor B \(Fermentation time\) B2 \(4 days\) B3 \(6 days\)](#) Mean A1 12.608 A2 12.44cB 13.37bB 14.97aB 13.596 A3 13.38cA 14.47bA 16.11aA 14.654 Mean 12.72 13.392 14.744 -- Note: Different uppercase letters in different columns and small [letters on the same row show very significant](#) different [\(P<0.05\)](#). Mean cellulase activity is in U/ml

Table 3. Mean protease activity (U/ml) of fermented palm kernel cake with *Bacillus subtilis*. Factor A [Factor B \(Fermentation time\)](#) (Inoculum Dose) [B1 \(2 days\) B2 \(4 days\) B3 \(6 days\)](#) A1 (3%) 5.85aB 6.34aB 5.36aC A2 (5%) 6.95bA 6.68abB 8.31aB A3 (7%) 7.21bAB 7.61bA 10.27aA Mean 6.67 6.87 7.98 Mean 6.38 7.31 8.36 J. World Poult. Res., 9(4): 211-216, 2019 Note: Different uppercase letters in different columns and small [letters on the same row show](#)

very **significant** different ($P < 0.05$). Mean Protease activity is in U/ml. CONCLUSION The high inoculum doses and the longer fermentation time can increase enzyme activity in fermented palm kernel cake. The 7% inoculum doses and 6-days fermentation time provided optimum results as indicated in mannanase activity (6.27 U/ml), cellulose activity (16.11 U/ml) and protease activity (10.27 U/ml). DECLARATIONS Acknowledgements This study was financially supported by funds provided by BOPTN of Andalas University, number 42/UN.16.17/PP.RGB/LPPM/2018, dated April, 23rd 2018. Author's contribution Mirnawati conducted the research, and prepared data. Gita Ciptaan did the field research and Ferawati performed statistical analysis. All authors checked and confirmed the final form of article Competing interests The authors have declared that no competing interest exists. Consent to publish All authors gave their informed consent prior to their inclusion in the study. REFERENCES Belma A, Zehra N and Yavuz B (2000). Determination of PHB Growth Quantities of Certain Bacillus Species Isolated From Soil. Journal of Biotechnology, Special Issue, pp. 24- 30. Available at: <http://www.biyotekder.hacettepe.edu.tr/dergi.html> Buckle KA, Edwards RA, Fleet GH and Wooton M (1987). Ilmu Pangan. UI Press, Jakarta. Central Bureau of Statistic (2015). West Sumatra in numbers. Central Bureau of Statistics, Padang. Cupp C and Enyard (2008). Sigma's non-specific protease activity assay - casein as a substrate. Journal of Visualized Experiments. 19:899. DOI: <http://10.3791/899>. Dada O, Kalil MS and Yusoff WMW (2012). Effect of Inoculum and substrate concentration in anaerobic fermentations of treated rice bran to acetone, butanol and ethanol. Bacteriology journal 2(4): 79-89. DOI: <http://10.3923/bj.2012.79.89> Darwis AA, Sailah I, Irawadi T T and Safriani (1995). Kajian Kondisi Fermentasi pada Produksi Selulase dari Limbah Kelapa Sawit (Tandan Kosong dan Sabut) oleh Neurospora sitophila. Jurnal Teknologi Industri Pertanian 5 (3) 199- 207. Available at: <https://repository.ipb.ac.id/jspui/bitstream/123456789/30559/1/F95SAF.pdf>. Daud MJ and Jarvis MC (1993). Mannan of oil palm kernel. Phytochemistry, 31: 463- 464. DOI: [https://doi.org/10.1016/0031-9422\(92\)90017-K](https://doi.org/10.1016/0031-9422(92)90017-K) Darwis AA and Sukara E (1990). Teknologi mikrobial. Departemen Pendidikan dan Kebudayaan. Direktorat Jendral Pendidikan Tinggi. Pusat Antar Universitas Bioteknologi, Institut Pertanian Bogor, pp. 100-165. Directorate General of Plantations (DGP) (2016). Indonesian Plantation Statistics Department of Agriculture, Jakarta Available at: <http://ditjenbun.pertanian.go.id/> Dwidjoseputro D (2010). Dasar-dasar Mikrobiologi, Edisi 14, Djambatan, Jakarta. Fardiaz S (1992). Teknologi Fermentasi. Jurusan Teknologi Pangan dan Gizi, Fakultas Pertanian, IPB, Bogor. Hooge D (2003). Bacillus spores may enhance broiler perform. Feed stuffs, 75: 1-5. Available at: https://www.calsporin.com/english01/info/img/index/feeds_tuffs.pdf Jiang Z, Wei Y, Li D, L Li, Chai P and Kusakabe I (2006). High- Level Production, Purification, and Characterization of A Thermostable-mannanase from the Newly Isolated Bacillus subtilis WY34. Carbohydrate Polymers, 66:88-96. DOI: <http://10.1016/j.carbpol.2006.02.030>. Mirnawati, Rizal Y, Marlida Y and Kompiani IP (2010). The role of humic acid in palm kernel cake

fermented by *Aspergillus niger* for poultry ration. *Pakistan Journal of Nutrition*, 9 (2): 182-185. DOI: <http://10.3923/pjn.2010.182.185>

Mirnawati, Kompiang IP and SA Latif (2012). Effect of substrat composition and inoculums dosage to improve quality of palm kernel cake fermented by *Aspergillus niger*. *Pakistan Journal of Nutrition*, 11(5): 434-438. DOI: <http://10.3923/pjn.2012.434.438>. Mirnawati, Djulardi A and Marlida Y (2013). Improving the quality of palm kernel cake fermented by *Eupenicillium javanicum* as poultry ration. *Pakistan Journal of Nutrion*, 12 (12): 1085-1088. DOI: <http://10.3923/pjn.2013.1085.1088>. Mirnawati, Djulardi A and Ciptaan G (2018). Utilization of fermented palm kernel cake with *Sclerotium rolfsii* in broiler ration. *International Journal of Poultry Science*. 17(7): 342-347. DOI: <http://10.3923/ijps.2018.342.347> Mirnawati, Ciptaan G and Ferawati (2017). The effect of Mananalytic fungi and humic acid dosage to improve the nutrient content and quality of fermented palm kernel cake. *International journal of Chem Tech Research*, 10(2): 56- 61. Available at: [http://www.sphinxsai.com/2017/ch_vol10_no2/1/\(56-61\)V10N2CT.pdf](http://www.sphinxsai.com/2017/ch_vol10_no2/1/(56-61)V10N2CT.pdf)

Mirnawati, Ciptaan G and Ferawati (2019). Improving the quality and nutrient content of palm kernel cake through fermentation with *Bacillus subtilis*. *International Journal of Animal and Veterinary Sciences*, 31(7). Available at: <http://www.lrrd.org/lrrd31/7/mirna31098.html>

Musaalbakri AM, Ariff A, Rosfarizan M and Ismail AKM (2005). Kinetics and modeling of red pigment fermentation by *Monascus purpures* FTC5391 in 2 liter stirred tank fermenter using glucose as a carbon source. *Journal Tropical Agriculture and Food Science*, 33: 277- 284. Available at: <http://psasir.upm.edu.my/id/eprint/48938/>

Rizal Y (2000). The respon of broilers to the substitution part of soybean meal for palm kernel cake in the diet. *Jurnal Peternakan dan Lingkungan*, 2: 15-20. Rizal Y, Nuraini, Mirnawati and Mahata ME (2013). Comparisons of nutrients contents and nutritional values of palm kernel cake fermented by using different fungi. *Pakistan Journal of Nutrition*, 12 (10): 943-948. DOI: <https://10.3923/pjn.2013.943.948>. Steel RGD and Torrie JH (1991). *Prinsip dan Prosedur Statistik suatu Pendekatan Biometrik*. PT. Gramedia Pustaka Utama, Jakarta.

Tafsin M (2007). Polisakarida mengandung manan dari bungkil inti sawit sebagai anti mikroba salmonella trypimurium pada ayam. *Media Peternakan*, 30: 139-146. Available at: <https://journal.ipb.ac.id/index.php/mediapeternakan/article/view/1003>.

Zulfatus S, Ika ISN and Abdullah (2008). Produksi Enzim Selulase oleh *Aspergillus niger* Menggunakan Substrat Jerami dengan Sistem Fermentasi Padat. (Online), Available at: http://eprints.undip.ac.id/13063/1/ARTIKEL_ILMIAH.pdf

Mirnawati et al., 2019 Mirnawati et al., 2019 Mirnawati et al., 2019 212 213 214 215 216