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Research Article

Ethyl Acetate Fraction Activities of *Myrmecodia tuberosa* Jack. in Anemic Mice

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Abstract

Background and Objective: Ethyl acetate fraction of *Myrmecodia tuberosa* Jack. increases phagocyte activity of macrophage and lymphocyte proliferation and also prevents cutaneous anaphylactic reactions. Based on that, this present research aim to investigate the effect of an acetate fraction from *Myrmecodia tuberosa* Jack. on numbers of erythrocyte, reticulocyte, hemoglobin content and hematocrit in mice. **Materials and Methods:** The research was conducted over 3 months and consisted of a positive control group and 3 groups treated with *Myrmecodia tuberosa* Jack. ethyl acetate fractions at 3 dosing levels. Anemia was induced in the mice using chloramphenicol 130 mg kg⁻¹ b.wt., for 14 days then for next 14 days daily oral doses of 40 mg kg⁻¹ b.wt., 63.2 mg kg⁻¹ b.wt., or 100 mg kg⁻¹ b.wt., of *Myrmecodia tuberosa* Jack. ethyl acetate fraction were administered to each group. Blood samples were taken on day 0, 14, 21 and 28 for analysis. Statistical analysis was conducted using two-way ANOVA then Duncan Multiple Range Test (DMRT). **Results:** About 40-63.2 mg kg⁻¹ b.wt. doses of *Myrmecodia tuberosa* Jack. ethyl acetate fraction significantly increased the erythrocyte, reticulocyte and hemoglobin count and hematocrit from the 14th day (p<0.01). **Conclusion:** Ethyl acetate fraction of *Myrmecodia tuberosa* Jack. could have potential as an anemia treatment.

Key words: Anemia, erythrocyte, hematocrit, hemoglobin, *Myrmecodia tuberosa* jack., reticulocyte

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Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Around 1,845 plants found in Indonesia are known to have been used as traditional medicine by different ethnic groups^{1,2}. The shrubby caudex forming epiphyte *Myrmecodia* sp. is commonly called *Sarang semut*, literally ant nest plant, has been used medicinally in Papua, Mentawai Islands and Borneo. This genus contains a number of species with putative medicinal properties including *Myrmecodia tuberosa*, *Myrmecodia pendants* and *Hydnophytum formicarum* (Rubiaceae)³. They are known to contain flavonoids, triterpenoid, tocopherol, polyphenol, glycoside, tannin as well as calcium, sodium, calcium, zinc, iron, phosphorus and magnesium^{4,5}.

It has been found that the ethyl acetate fraction of *Myrmecodia tuberosa* can increase the phagocytic activity of macrophages and increase lymphocyte production *in vitro*⁶. These effects are thought to be related to the activity of phenol and flavonoid compounds. There is hope that by increasing lymphocyte cell proliferation these compounds could have anti-cancer properties⁷ and could prevent active cutaneous nephrotoxic reactions⁸. The ethanol extract of *Myrmecodia tuberosa* has been found to increase Sprague Dawley (SD) mouse TCD4+ and TCD8+ *in vivo* after doxorubicin treatment⁹. *Myrmecodia tuberosa* flavonoids appear to have strong anti-inflammatory properties¹⁰. Terpenoid from *Myrmecodia tuberosa* has been found to have anti-cancer properties especially for human cervical cancer¹¹. Flavonoids from *Myrmecodia tuberosa* killed a significant percentage of tongue cancer SP-C1 cells¹². Also, a water extract of *Myrmecodia tuberosa* appears to cure diarrhea and improve bowel function¹³.

When mature mammalian erythrocytes emerge from bone marrow they live about 120 days until disintegration and death. Dead erythrocytes are replaced by new cells which are produced by the bone marrow. White blood cells, unlike erythrocytes contain a nucleus and move independently. These are produced in the bone marrow and lymph nodes and play a role in eradicating disease¹⁴.

Blood count is one indication of health status. Blood transports nutrients, oxygen, carbon dioxide, metabolites, hormones, antibodies and is essential in maintaining fluid balance and body pH¹⁵.

Anemia results from lack of total blood or erythrocytes in the blood which hinders the transport of oxygen around the body. Erythrocytes contain the iron-containing complex protein hemoglobin. Anemia occurs when the hemoglobin level drops below 12 g dL⁻¹ b.wt., for a woman or 14 g dL⁻¹ b.wt., for a man. Low hematocrit value and

reticulocyte count can also indicate the type of anemia present and the status of bone marrow, where erythrocytes are produced^{16,17}.

Anemia occurs frequently because of malnutrition leading to deficiency in iron, folic acid or B₁₂ but it can also be a result of damage to the stomach or compromised renal function leading to reduced erythropoietin production and infection. Anemia can also be a result of excessive breakdown and loss of erythrocytes due to heavy menstrual bleeding, childbirth, hemolysis or use of sustenance that irritate the stomach^{18,19}.

While anemia is a particular problem in isolated areas, low availability and lack of affordability put modern anemia medicines out of reach of those who most need it. Sometime the problem may be due to nutritional deficiencies but often what is needed is a way for the bone marrow to be stimulated to produce more erythrocytes. If components found in readily available and easy to cultivate plants can be found to achieve this aim then this could provide a solution to this problem.

Myrmecodia tuberosa Jack. already used traditionally to treat anemia but no research has been conducted to determine its effectiveness or appropriate dose or duration of treatment. As flavonoids are thought to be the active ingredient in this plant in stimulating erythrocyte production, in this present study these were extracted from *Myrmecodia tuberosa* Jack. using ethyl acetate and their effect on anemic mice investigated. Parameters measured were erythrocyte and reticulocyte counts, hemoglobin level and hematocrit value.

MATERIALS AND METHODS

Time and place: The research was conducted in July-September, 2017 at KOPERTIS Laboratory Region X, Pharmacy Research Laboratory, Faculty of Pharmacy Universitas Andalas and Serology-Immunology Laboratory of Faculty of Pharmacy Universitas Andalas.

Materials: The materials on this research consist of *Myrmecodia tuberosa* Jack. Figure 1, ethyl acetate 1%, aquadest, ethanol 96%, Tween-80 0.1%, Carboxymethylcellulose (CMC) 0.1%, Drabkins Reagent (Catalog number: D5941 Sigma), Hayem solution (Catalog number: MFCD01866932 Sigma), cresyl blue brilliant 1% and chloramphenicol 200 mg mL⁻¹.

Equipment: Animal scales, maceration bottle, mortar, stamper, mice cage, measuring glass, sonde needles, thin-layer chromatography (TLC) plate, hematocrit pipette, hemoglobin pipette, Hettich centrifuge, Uv-Visible (BIO-RADx



Fig. 1: A fresh "ant nest" tuber *Myrmecodia tuberosa* Jack

Mark) spectrophotometer, erythrocyte pipette, hemocytometer and microscope (ZEISS).

Animal experimentation: Twenty mice (*Mus musculus*, Swiss webster strain) 2-3 months-old with body mass 20-30 g from Pharmacology Laboratory, Faculty of Pharmacy Universitas Andalas were used. About 7 days were allowed for acclimatized and observation before treatment began.

Extraction and fractionation *Myrmecodia tuberosa* Jack: About 4 kg of fresh *Myrmecodia tuberosa* Jack. were sliced into 2-3 mm slices then dried in a greenhouse for 3 days then in a 50°C oven for 3 days. These were then blended to produce 400 g of powder which was placed in a dark macerator bottle with 4 L of 70% ethanol solvent, soaked for three days, stirring occasionally. The mixture was then filtered with filter paper four times until clear. The residue was then evaporated *in vacuo* with a rotatory evaporator until a thick extract was obtained^{20,21}.

This extract was dissolved in an equal volume of aquades and ethyl acetate solvent. The ethyl acetate fraction was pipetted off then evaporated until a viscous fraction remained.

Characterization of the viscous fraction: The viscous fraction of ethyl acetate was examined organoleptically and a rendement test conducted. The moisture and ash content was determined as was the TLC profile.

Thin layer liquid chromatography: A thin layer liquid chromatography profile of the ethyl acetate fraction was conducted using an eluent made from a mixture of butanol: acetate acid: water (2:0.5:2.5). The flavonol quercetin was used as a comparison.

Treatment of mice: About 130 mg kg⁻¹ b.wt., dose of chloramphenicol was given to each mouse every day for 14 days. Chloramphenicol suppresses the proliferation and differentiation of erythrocytes reducing the erythrocyte count in the blood producing anemia²². The anemic mice were divided into four groups. The positive control group was orally dosed with a physiological saline solution and the second, third and fourth groups were given an oral daily 40, 63.2 and 100 mg kg⁻¹ b.wt., dose of the ethyl acetate fraction respectively.

Erythrocyte count: A pipette rinsed with Hayem solution, the tail of the mouse was cut off and the wound cleaned with a cotton swab. 0.5 µL of the blood from the mouse was suctioned into the pipette and the tip of the pipette cleaned with tissue. Sufficient Hayem solution was pipetted up after the blood to make a total of 101 µL. The filled pipette was shaken for 3 min, two drops discarded then the tip placed on a glass slide and covered with a coverslip. After 2-3 min for the erythrocytes to settle a count was made under a microscope at 400x enlargement^{19,23}.

Reticulocyte count: Blood and brilliant cresyl blue dye were mixed with ratio 1:1 in a tube and set aside for 15 min for the dye to be absorbed by the blood cells. About 1-2 drops were dried on a slide then examined under a microscope at 100x. Reticulocytes contain blue granules/filaments while mature erythrocytes appear as clear light blue disks. The ratio of reticulocytes to 1000 erythrocytes was counted^{19,23}.

Hemoglobin level: Five milliliter Drabkin solution was mixed with 20 µL blood and shaken in a tube until well mixed then set aside at room temperature for 3 min. Hemoglobin Level was determined using a spectrophotometer to measure absorbance at 546 nm^{19,23}.

Hematocrit level: Mouse blood was pipetted into a microcapillary pipette until $\frac{3}{4}$ full and one tip stopped with wax. The tube was centrifuged (microhematocrit centrifuge) at 16000 rpm for 5 min. The Hematocrit level was measured by comparing the height of the solid fraction with the height of the solution in the microcapillary pipette.

Data analysis: Correlations of these blood parameters with ethyl acetate dose were measured using two-way ANOVA. If significant correlations were found at the p<0.05 level these were further tested using DMRT (with IBM SPSS V20.0).

RESULTS AND DISCUSSION

The ethyl acetate fraction was viscous, aromatic, black-brown and bitter. It contained 5.59% rendement, 11.44% moisture content and 6.24% ash. Results of the TLC indicate that only one major flavonol is present and it is, in fact, quercetin. The fraction had a retardation factor Rf of 0.78. The TLC profile can be seen in Fig. 2.

Erythrocyte counts (million μL^{-1}) for 14 days of ethyl acetate fraction administration is shown in Table 1. The result of statistical analysis (Table 2) suggested that the erythrocyte count after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack. increased significantly ($p < 0.01$) for all doses and durations of treatment. The increase in erythrocytes after 63.2 and 100 mg kg^{-1} b.wt., doses were not significantly different at the $p < 0.01$ level. The DMRT Test (Table 3) indicated that increasing dose size and duration of treatment significantly increases erythrocyte count ($p < 0.01$), however, the difference between 63.2 and 100 mg kg^{-1} b.wt., doses was not significant.

Erythrocytes are the most numerous blood cells. There are many more erythrocytes compared to leukocytes and

platelets. After emerging from the bone marrow where they are produced they live about 120 days before disintegrating and being replaced by new cells^{14,24}. Erythrocytes contain hemoglobin which allows red blood cells to carry oxygen from the lungs and deliver it throughout the body tissues²⁵. Anemia, lack of the ability of the blood to carry oxygen,

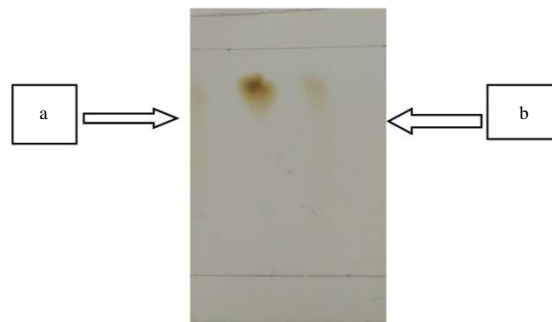


Fig. 2: TLC profile of the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. under UV light (254 nm) using an eluent mixture of butanol: acetate acid: water (2:0.5:2.5) a: Quercetin standard, b: Ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Table 1: Erythrocyte cells count in mice with anemia induced by 14 days of chloramphenicol and subsequent dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack. at different doses

Doses	Amount of erythrocyte (millions μL^{-1})			Average \pm SD
	Day 14	Day 21	Day 28	
Positive control	4.39 \pm 0.19	4.83 \pm 0.20	5.25 \pm 0.14	4.82 \pm 0.40
Dose 40 mg kg^{-1} b.wt.	4.39 \pm 0.19	5.18 \pm 0.26	5.58 \pm 0.20	5.04 \pm 0.56
Dose 63.2 mg kg^{-1} b.wt.	4.41 \pm 0.13	5.59 \pm 0.36	5.86 \pm 0.26	5.29 \pm 0.70
Dose 100 mg kg^{-1} b.wt.	4.45 \pm 0.15	5.61 \pm 0.08	5.99 \pm 0.09	5.35 \pm 0.69
Average \pm SD	4.41 \pm 0.15	5.30 \pm 0.40	5.67 \pm 0.62	

Table 2: Two-way ANOVA analysis of erythrocyte count after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Sources	Type III sum of squares	df	Mean square	F-value	Significant
Doses	2.638	3	0.879	21.517	0.000
Duration	16.927	2	8.464	207.115	0.000
Doses and duration	1.099	6	0.183	4.483	0.001
Total	1599.179	60			

df: Degree of freedom

Table 3: DMRT analysis of erythrocyte count after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Treatments	N	Subset for alpha = 0.05		
		1	2	3
Doses				
Positive control	20	4.8213		
40 mg kg^{-1} b.wt.	20		5.0447	
63.2 mg kg^{-1} b.wt.	20			5.2880
100 mg kg^{-1} b.wt.	20			5.3500
Significant		1.0000	1.0000	0.4050
Duration				
14th day	20	4.4050		
21st day	20		5.3040	
28th day	20			5.6690
Significant		1.0000	1.0000	1.0000

occurs in mammals whenever hemoglobin level drops below 12 g dL⁻¹ b.wt., for female and 14 g dL⁻¹ b.wt., for male. Anemic individuals also have lower hematocrit levels and reticulocyte counts. Hematocrit levels are useful to diagnose the type of anemia and reticulocyte counts indicate the condition of the bone marrow where they are produced.

The used of chloramphenicol in this research served as an anemia inducer administered for 14 consecutive days. Chloramphenicol works to suppress the bone marrow so that it inhibits proliferation and differentiation. Thus, the formation of erythrocyte components can be inhibited and cause anemia. Anemia caused by chloramphenicol is classified as aplastic anemia. Anemia aplastic is a deficiency of erythrocytes, reticulocytes, hemoglobin and hematocrit as a result of reduction of erythroblast cells being produced in the bone marrow^{22,26}.

Erythrocytes develop from hemocytoblast cells. New hemocytoblasts will continuously form from bone marrow stem cells. Hemocytoblasts form basophilic erythroblasts which begin to synthesize hemoglobin and then erythroblast turns into polychromatophilic erythroblasts, then the nuclei of these cells grow smaller and the cells produce hemoglobin and become normoblast. After the cytoplasm of the normoblast is filled with hemoglobin, the nuclei disappear and endoplasmic reticulum are reabsorbed by the cells. These cells are now called reticulocytes because they still contain a few basophilic endoplasmic reticula which stays with the hemoglobin inside the cytoplasm. The endoplasmic reticulum undergoes capillary diapedesis, slipping out of the reticulocytes through membrane pores. After the reticulum is all reabsorbed, cells become matured erythrocytes¹⁶.

The reticulocyte count for 14 days of ethyl acetate fraction administration is shown in Table 4. The increase is highly significant (p<0.01). The increase due to 40 mg kg⁻¹ b.wt., and 63.2 and 100 mg kg⁻¹ b.wt., was highly significantly different (p<0.01).

The effect of ethyl acetate fraction dose and duration of treatment of reticulocyte is shown in Table 5 and 6. The ethyl acetate fraction dose showed a similar relationship with the reticulocyte count as it does with the erythrocyte count. This is to be expected as the reticulocytes develop into erythrocytes so an increase in one implies an increase in the other.

The increase in reticulocyte count suggested that, as expected, chloramphenicol only caused reversible suppression of the bone marrow function and did not permanently damage its ability to produce erythropoietin²⁷. On the contrary, the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. appears to stimulate reticulocyte production in the bone marrow²⁶. An increase in the number of reticulocytes in peripheral blood indicates increased production of erythrocytes in the bone marrow. A low reticulocyte count would indicate bone marrow hypofunction or aplastic anemia^{28,29}.

The average content of hemoglobin (g dL⁻¹ b.wt.) for 14 days of ethyl acetate fraction administration is shown in Table 7. ANOVA analysis indicated a significant relationship between dose and duration on the hemoglobin level (p<0.05) (Table 8). Subsequent DMRT results (Table 9) showed while neither the 40 or 63.2 mg kg⁻¹ b.wt., dose resulted in hemoglobin levels significantly higher than the positive control, the 100 mg kg⁻¹ b.wt., dose did result in a significant increase (p<0.05).

Table 4: Reticulocyte count in mice with anemia induced by 14 days of chloramphenicol and subsequent dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack. at different doses

Doses	Amount of reticulocyte (millions μL^{-1})			Average \pm SD
	Day 14	Day 21	Day 28	
Positive control	0.42 \pm 0.04	0.68 \pm 0.08	0.78 \pm 0.04	0.63 \pm 0.17
Dose 40 mg kg ⁻¹ b.wt.	0.48 \pm 0.08	0.76 \pm 0.11	0.86 \pm 0.09	0.70 \pm 0.19
Dose 63.2 mg kg ⁻¹ b.wt.	0.44 \pm 0.05	0.78 \pm 0.08	0.96 \pm 0.11	0.73 \pm 0.24
Dose 100 mg kg ⁻¹ b.wt.	0.42 \pm 0.08	1.02 \pm 0.15	1.38 \pm 0.13	0.94 \pm 0.43
Average \pm SD	0.44 \pm 0.07	0.81 \pm 0.17	0.99 \pm 0.25	

Table 5: Two-way ANOVA analysis of reticulocyte count after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Sources	Type III sum of squares	df	Mean square	F	Significant
Doses	0.815	3	0.272	30.191	0.000
Duration	3.194	2	1.597	177.463	0.000
Doses and Duration	0.588	6	0.098	10.895	0.000
Total	38.630	60			

df: Degree of Freedom

Table 6: DMRT analysis of reticulocyte count after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Treatments	N	Subset for alpha = 0.05		
		1	2	3
Doses				
Positive control	20	0.6267		
40 mg kg ⁻¹ b.wt.	20		0.7000	
63.2 mg kg ⁻¹ b.wt.	20		0.7267	
100 mg kg ⁻¹ b.wt.	20			0.9400
Significant		1.0000	0.4450	1.0000
Duration				
14th day	20	0.4400		
21st day	20		0.8100	
28th day	20			0.9950
Significant		1.0000	1.0000	1.0000

Table 7: Hemoglobin levels in mice with anemia induced by 14 days of chloramphenicol and subsequent dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack. at different doses

Doses	Content of hemoglobin (g dL ⁻¹)			Average ±SD
	Day 14	Day 21	Day 28	
Positive control	11.93 ± 1.13	14.37 ± 0.65	15.67 ± 0.69	13.99 ± 1.79
Dose 40 mg kg ⁻¹ b.wt.	11.98 ± 0.55	14.96 ± 0.58	15.77 ± 0.65	14.24 ± 1.77
Dose 63.2 mg kg ⁻¹ b.wt.	12.10 ± 0.59	15.02 ± 1.47	15.91 ± 1.78	14.34 ± 2.12
Dose 100 mg kg ⁻¹ b.wt.	12.14 ± 0.33	17.06 ± 1.40	18.20 ± 1.81	15.80 ± 2.99
Average ±SD	12.04 ± 0.66	15.35 ± 1.46	16.39 ± 1.65	

Table 8: Two-way ANOVA analysis of hemoglobin levels after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Sources	Type III sum of squares	df	Mean square	F	Significant
Doses	30.103	3	10.034	8.403	0.000
Duration	206.559	2	103.279	86.484	0.000
Doses and duration	12.834	6	2.139	1.791	0.121
Total	13082.989	60			

df: Degree of freedom

Table 9: DMRT analysis of hemoglobin levels after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Treatments	N	Subset for alpha = 0.05		
		1	2	3
Positive control	20	13.9887		
40 mg kg ⁻¹ b.wt.	20	14.2380		
63.2 mg kg ⁻¹ b.wt.	20	14.3440		
100 mg kg ⁻¹ b.wt.	20		15.7987	
Significant		0.4070	1.0000	
Duration				
14th day	20	12.0375		
21st day	20		15.3515	
28th day	20			16.3880
Significant		1.0000	1.0000	1.0000

Hemoglobin carries iron ions called heme and globulin protein. There are around 300 hemoglobin in one erythrocyte. Hemoglobin carries oxygen from the lungs to other parts of the body and brings carbon dioxide back to the lungs where it is exhaled¹⁴. So the increase in hemoglobin due to the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. indicates an improved ability of the blood to transport oxygen.

Hematocrit values measured for 14 days of ethyl acetate fraction administration are shown in Table 10. There was a significant relationship between ethyl acetate fraction dose and duration of treatment (Table 11, 12) and hematocrit value (p<0.05). Meanwhile, the interaction between doses of treatment and days of monitoring indicates there was no significant effect on the amount of hematocrit content (p>0.05). Thus, the effect caused by an ethyl acetate fraction

Table 10: Hematocrit value in mice with anemia induced by 14 days of chloramphenicol and subsequent dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack. at different doses

Doses	Value of hematocrit (%)			Average±SD
	Day 14	Day 21	Day 28	
Positive control	41.9±1.75	44.2±1.48	45.6±1.48	43.9±2.09
Dose 40 mg kg ⁻¹ b.wt.	43.8±2.49	45.0±1.58	46.9±2.22	45.2±2.37
Dose 63.2 mg kg ⁻¹ b.wt.	43.2±2.59	45.2±1.95	47.4±1.14	45.3±2.56
Dose 100 mg kg ⁻¹ b.wt.	43.5±2.57	46.7±2.73	49.2±2.92	46.5±3.50
Average±SD	43.1±2.30	45.3±2.06	45.2±2.77	

Table 11: Two-way ANOVA analysis of hematocrit values after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Sources	Type III sum of squares	df	Mean square	F-value	Significant
Doses	49.483	3	16.494	3.622	0.019
Duration	174.408	2	87.204	19.148	0.000
Doses and duration	10.692	6	1.782	0.391	0.881
Total	123126.000	60			

df: Degree of freedom

Table 12: DMRT analysis of hematocrit values after dosing with ethyl acetate fraction of *Myrmecodia tuberosa* Jack

Treatments	N	Subset for alpha = 0.05		
		1	2	3
Doses				
The positive control	20	43.900		
40 mg kg ⁻¹ b.wt.	20	45.233	45.233	
63.2 mg kg ⁻¹ b.wt.	20	45.267	45.267	
100 mg kg ⁻¹ b.wt.	20		46.467	
Significant		0.103	0.141	
Duration				
14th day	20	43.100		
21st day	20		45.275	
28th day	20			47.275
Significant		1.000	1.000	1.000

of *Myrmecodia tuberosa* Jack. on the value of hematocrit value was highly significant.

Furthermore, this research can be continued to determine the activity of the active compounds in the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. by observing the cytokine and erythropoietin (EPO) production of cells under hypoxic conditions along with interleukin-1 (IL-1) and interleukin-9 (IL-9). These cytokine compounds are responsible for the proliferation and differentiation of stem cells into pronormoblasts then into erythrocytes. It is expected that the active compounds present in *Myrmecodia tuberosa* Jack. can support some stages of the process of proliferation and differentiation in the process of erythrocyte formation and not affect other cells.

CONCLUSION

The conclusion of this research are the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. at doses of 40, 63.2 and 100 mg kg⁻¹ b.wt., can increase the formation of erythrocytes

in anemic mice. The higher the doses of ethyl acetate fraction *Myrmecodia tuberosa* Jack., faster erythrocytes are produced. This suggested that *Myrmecodia tuberosa* Jack. has potential as an economic and effective source of treatment for some types of anemia.

SIGNIFICANCE STATEMENT

This study discover the ethyl acetate fraction of *Myrmecodia tuberosa* Jack. was able to increase the amount of erythrocyte, reticulocyte, the content of hemoglobin and value of hematocrit in mice that can be beneficial as an effective treatment for many anemias. *Myrmecodia tuberosa* Jack. grows abundantly in the Mentawai Islands. In isolated tropical areas, anemia due to hepatitis, pregnancies and childbirth, malaria and kidney disorders are significant problems. These are all anemias that could well be treated using an extract of *Myrmecodia tuberosa* Jack. This study will help the researcher to uncover the critical areas of effectiveness of *Myrmecodia*

tuberosa Jack. against anemia. This plant could well become an economic and easily available treatment.

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