

III - A - 2 - a - 1 - a



Proceeding

International Seminar on Food & Agricultural Science

17 February 2010 Bukittinggi, Indor

*"Improving the quality of life
through food and agricultural sciences"*

EDITORS

- Novizar Nazir - Universitas Andalas, Indonesia
- Aisman - Universitas Andalas, Indonesia
- Nazaruddin Ramli - Universiti Kebangsaan Malaysia
- Wan Aida Wan Mustapha - Universiti Kebangsaan Malaysia
- Ayub Mohd Yatim - Universiti Kebangsaan Malaysia
- Mohamad Kassim - Universiti Kebangsaan Malaysia
- A. Ploeger - Kassel University Germany



PROCEEDING

INTERNATIONAL SEMINAR ON FOOD AND AGRICULTURAL SCIENCES-ISFAS2010

16-17 FEBRUARY 2010

HILL HOTEL AND CONVENTION, BUKITTINGGI-INDONESIA

Improving the quality of life through food and agricultural sciences

Editors:

Novizar Nazir (*Universitas Andalas, Indonesia*)
Aisman (*Universitas Andalas, Indonesia*)
Nazaruddin Ramli (*Universiti Kebangsaan Malaysia*)
Wan Aida Wan Mustapha (*Universiti Kebangsaan Malaysia*)
Ayub Mohd Yatim (*Universiti Kebangsaan Malaysia*)
Mohamad Kassim (*Universiti Kebangsaan Malaysia*)
A. Ploeger (*Kassel University-Germany*)

First published in 2010 by:

AgriTech Press

Faculty of Agricultural Technology - University of Andalas
Gedung Fateta Level 2- Kampus Unand Limau Manis
Padang, Indonesia 25163- Telp. +62 751 72772. Fax. +62 751 72702
<http://www.fateta.unand.ac.id>

All right reserved. This book, or part of thereof, may not be reproduced in any form by any means, electronic or mechanical, including photocopying, recording or any information storage and retrieval system know or to be invented, without written permission from the publisher.

Printed and bound in Malaysia by Pusat Penerbitan dan Percetakan
Universiti Kebangsaan Malaysia, 43000 UKM Bangi-Selangor D.E-Malaysia

Cover design and layout:
Rahmat Hidayat

© 2010 AgriTech Press
Faculty of Agricultural Technology - University of Andalas
ISBN 978-602-96301-0-7

		RESIDUAL EFFECT OF PHOSPHORUS AND ORGANIC MATTER IN NORTH SUMATRA
30	Rachmi Yusuf and Ali Jamil	APPLICATION OF PLANT EXTRACT ON KANGKUNG (<i>IPOMEA REPTANS</i> L) CROP GROWTH IN RIAU
32	Ida Nur Istina, Parlin H Sinaga, Dorlan S and Ali Jamil	FEASIBILITY STUDY FOR COCOA CROP DEVELOPMENT IN RIAU PROVINCE
33	Irawati Chaniago and Irfan Suliansyah	IN SEARCH FOR LOCAL WISDOMS IN WEED MANAGEMENT IN RICE CULTIVATION.
36	Yulmafismawita, Gusnidar, and Amrizal Saidi	ROLE OF ORGANIC MATTER IN SITU FOR AGGREGATE STABILITY IMPROVEMENT OF ULTISOL IN WEST SUMATRA AND CHILLI (<i>CAPSICUM ANNUM</i>) PRODUCTION
40	Hidriyanti	THE ROLES OF DIFFERENT INTERCROPPING PLANTS IN MANAGING CAULIFLOWER PESTS AND THEIR PARASITIDS
44	Marini Wijayanti, Dade Jubacdah , & Ria Puspitasari	SPIRULINA PRODUCTION IN FERTILIZER MEDIUM COMBINED TOFU AND LATEX LIQUID WASTES".
47	Anthony Hamzah ¹⁾ , Wuri Prameswari ²⁾ dan Jamsari ³⁾	DIVERSITAS BAKTERI PELARUT FOSFAT PADA RHIZOSFER DAN RHIZOPLAN TANAMAN TOLERAN LAHAN GAMBUT PROPINSI RIAU
50	Muhammad Affan Fajar Falah , Arita Dewi Nugrahini, Shyntia Atica Putri, Agustinus Suryandono and Damawan Ari Nagroho	MAINTAINING FRESHNESS QUALITY OF FRESH-CUT ROSE (<i>ROSA SP</i>) FLOWER THROUGH CONTROLLED ENVIRONMENT AND SUCROSE PULSE TREATMENT.
58	Trizelia	PROSPECT OF ENTOMOPATOGENIC FUNGUS <i>BEAUVERIA BASSIANA</i> TO CONTROL <i>CROCIDOLomia PAVONANA</i> F (LEPIDOPTERA: CRAMBIDAE) PEST
57	Melly Novita, Siti Hajar Ahmad, Phebe Ding and M. Ridzwan Abd Halim	PHYSICO-CHEMICAL CHANGES OF RED PITAYA (<i>HYLOCEREUS POLYRHIZUS</i>) AS AFFECTED BY HARVEST DATE AND 1-MCP TREATMENT
62	Jamsari , Jamsu Trisno, Ade Noverta , Irfan Sultanyah, Ishak Manti , Nasrun	GENOME ANALYSIS OF GEMINIVIRUS IN PEPPER CULTIVATION FROM WEST SUMATERA
66	Jamsar, Anthony Hamzah, Yusniwati., Neswati, Wuri Prameswari, Selvi Elvia, Vovien and Cut Andesti.	SPECIES IDENTIFICATION OF BACTERIA FROM EXTREME HABITAT USING 16S-RRNA GENE SEQUENCES.
70	Ediwirman and Yusmanidar Arifin	MOLECULAR CHARACTERIZATION OF DURIO LOCAL FROM WEST SUMATERA BASED ON PCR-RAPD
74	Ichwana	SOIL HUMIDITY DYNAMICS ON BIOPORE ABSORPTION HOLE FOR RESIDENTIAL AREAL AND CULTIVATED LAND

GENOME ANALYSIS OF GEMINIVIRUS IN PEPPER CULTIVATION FROM WEST SUMATERA

Jamsari¹⁾, Jumsu Trisno²⁾, Ade Noverta¹⁾, Irfan Suliansyah¹⁾, Ishak Marti³⁾, Nasir

1. Lab. of Biotechnology and Plant Breeding, Fac. of Agriculture Andalas University

2. Department of Plant Pathology, Fac. of Agriculture Andalas University

3. Agency of Agricultural Technology Study and Application in West Sumatera.

4. Field Research Laining-Solok

* Corresponding author. ajamsari@yahoo.com

Abstract

Analysis of DNA components that composed genome of geminivirus is a prerequisite in order for effectively controlling this pathogen. For that reason primer walking strategy was applied. Forty-six primers in total have been synthesized and tested for the identification of DNA components in the geminivirus genome. All the designed primers have been tested in six different isolates. The result successfully identified of DNA-A, and satellite DNA but no DNA-B could be amplified from geminivirus from West Sumatera. Sequencing method was performed bidirectional in order to elucidate the genome structure. After editing the sequence data, currently 1464 bases was collected and analyzed. The length is approximately 54% from the total of gemini virus genome. Analysis of β -component (satellite DNA) was successfully amplified from isolate TD-21. The sequence composed of 1347 bases. In conclusion some facet could be mentioned that genome structure of geminivirus from West Sumatera is monopartite as proved by the absence of DNA component and the presence of β -component. About 54% of the DNA-A component was successfully sequenced and analyzed and the other 46% is still in progress.

Keywords

Gemini Virus, DNA-A, DNA-B, satellite DNA, Confg.

INTRODUCTION

Geminivirus is one of the main constraints in pepper cultivation. It was identified in the first time at 1999 in West Java [4], and known as a causing agent of Pepper Yellow Leaf Curl Disease. Since 2000 the disease has spread in all Java Island and was regarded as epidemic in all pepper cultivation [9]. Field identification of the disease symptom caused by geminivirus is compounded by the symptom caused by other viruses which are mosaic, stunt, yellow leaf, curly and malformation. The symptom mainly could be observed during flowering stage and very rare during transplantation period. Currently no effective strategy has been reported. Resistant cultivar should be an effective and economic strategy [1]. However, no resistant cultivar for this purpose is currently available so far. With the advent of DNA engineering technology it is possible to create transgenic crop that could be resistant against geminivirus

PCR Reaction and Sequence Analysis.

A universal primer pair as described by Roum was used as starting point of sequence analysis. The primers were PAL1978 (5'-GCACTGACAGCCGATTCGTTTCYCCNGT-3') and PAR1975 (5'-GATTG-TGCAGTTDATTCTTTCRTTCATCCAT-3'). A fragment of 1600 bp in length should be produced using this primer pair. Amplification was done in 25 μ l total volume reaction, containing 1 μ l of DNA template, 1 μ l of combined primer pair (5 pmol/l), and 1 bead of RTG PCR bead (Ge-Healthcare-UK). PCR reaction was done in a Biometra Thermocycler by applying the denaturation (94°C in 4 minutes), 30 core cycles

Material and Method

We collected 50 infected plants from six different regions in West Sumatera. Every region represents geographical conditions (lowland, medium land and highland). Virus DNA isolation was done as described by many authors [2, 3, 6]. Briefly leaf of infected plants prepared in 1.5 ml microcentrifuge tube. One microvolume of buffer containing 0.1 M Tris (pH 8.3), 0.05 M EDTA and 1% SDS was added inside. Leaf sample grinded well with plastic pestle before incubate for 5 minutes at 65°C. Three point five microliter of β -mercaptoethanol was added and the tube successively stored for 30 minutes on ice. After that, the tube was centrifuged for 15 minutes in 10,000 rpm. The supernatant was done two times extraction with Chloroform (1:1). The supernatant was finally saturated in TE. The supernatant was transferred into the new sterile tube and precipitated with 2.5 ml of 100% ethanol. The pellet was dried and resuspended in 25 μ l of sterile H₂O.

Isolates Collection and DNA Isolation

We collected 50 infected plants from six different regions in West Sumatera. Every region represents geographical conditions (lowland, medium land and highland). Virus DNA isolation was done as described by many authors [2, 3, 6]. Briefly leaf of infected plants prepared in 1.5 ml microcentrifuge tube. One microvolume of buffer containing 0.1 M Tris (pH 8.3), 0.05 M EDTA and 1% SDS was added inside. Leaf sample grinded well with plastic pestle before incubate for 5 minutes at 65°C. Three point five microliter of β -mercaptoethanol was added and the tube successively stored for 30 minutes on ice. After that, the tube was centrifuged for 15 minutes in 10,000 rpm. The supernatant was done two times extraction with Chloroform (1:1). The supernatant was finally saturated in TE. The supernatant was transferred into the new sterile tube and precipitated with 2.5 ml of 100% ethanol. The pellet was dried and resuspended in 25 μ l of sterile H₂O.

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

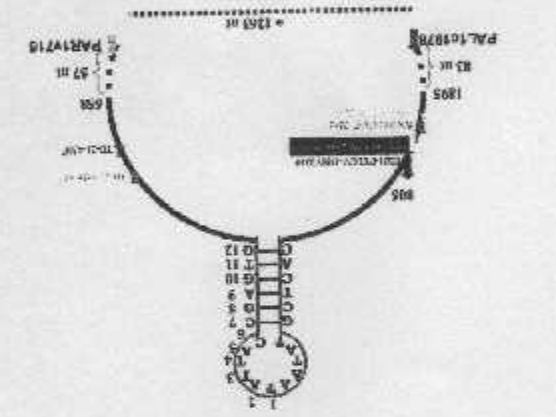
The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

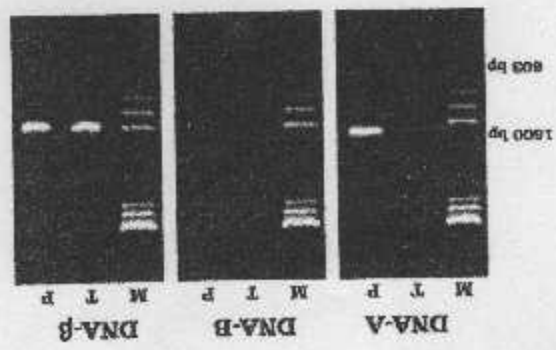
CTCTCCCTATGATGGCTGATGGCTCTG
GGCAGCCCAAGGGCTTCATCAGATTTCAGCA
GCCATTGTTCGATTCTCTGGAACACTTGTGA
ACACGACGACGATATGGTGAACAAATGTAT
CTACAGGTTAGCAAAATATTTATGAAATTAAC
ACTAATATTTGGTACTGTAGAACATAAGTCTTC
GGAGCAAGCTCTTAAATCAATCGAAGACATCG
GACTTACTCCACAATTAAGGGCCCTGGCGTAAA
CATCGTTACTGGTGGAGACCACTCGTGCAGA
TCGGCCATCTATGAAATTCCTCCATTGGTG
GTATCACCGGCTTTTGTCCATGTACCGCTGACAT

Figure 2. Schematic diagram of DNA-A genome of TD-21 isolate. Predicted gap might be 1.3 kb (1263 bases). Markers developed for primer walking are designated with arrowhead.



Mapping of whole genome structure of Gemintivirus from isolate TD-21 was performed with primer walking strategy. The additional primers were designed in the internal region of existing previous sequence data. Nucleotide sequence data indicated that 54% of TD-21 genome was successfully elucidated. This was achieved by applying both primers PAR1975 and PAR1978 in sequencing of 1600 bp fragment.

Figure 1. Electrophoresis performance of some primer pairs used to amplify DNA-A, DNA-B and satellite DNA (DNA-β) in TD-21 (T) and PSS-14 (P).



Whole genome sequencing and analysis.

The PCR product was purified using the High Pure PCR Purification Kit (Roche Diagnostics, Mannheim, Germany). The purified PCR product was digested with *Hpa*I and *Xba*I. The digested DNA was ligated into the *Hpa*I-*Xba*I digested pUC19 vector using the ligation kit (Roche Diagnostics). The recombinant plasmid was transformed into *E. coli* DH5α cells. The plasmid was extracted from the transformed cells using the High Pure Plasmid Purification Kit (Roche Diagnostics). The plasmid was sequenced using the BigDye 3.1 sequencing kit (Applied Biosystems, Foster City, CA, USA) on an ABI 3130XL DNA sequencer (Applied Biosystems).

TTAGCTACG
CCGAGCTTAGCTGCGCAGCTTATATACC
GGTGGCCGAGCGGTTCTCTGGTGTCCAGATGGTG

genome can be seen in Figure 3.
(Figure 2). Partial detail nucleotide sequences of DNA-A
meant 1.3 kb should be obtained using primer walking
during total genome of TD-21 genome was 2,7 kb, it
259 bases. The contig spanned along 1465 bases. Consi-
both primers showed an overlapping region in length of
primer. Contig assembling of nucleotide sequence from
quality of sequence data in average 862 bases for each
Primer PAR1c1975 and PAL1c1978 produced high
shown in legend.

TD-21 isolate spanned 1464 bases in length.
Iron, nonnucleotide and IR region was

(IR), where \square predicted as first base in the genome map.
= inverted region

\square = Iron 1 (688-694)
 \square = Iron 2 (715-728)
 \square = TATA Box (729-734)
 \square = nonnucleotide (800-812)

CGGCATTAAT
ACTGTTGTCATCTGCAAGGCCACATTTGTCCTC
GGTTCCGATGTCACAAGAGGTAATGGTTTGA
CGACATGACATTAACATGACTGGGAAGGCCCTT
TGAAGGACCTTGAAGTCCATCTTTGAACAG
GGTTCGAAGGAGCAGTGTATGTTCCGGGGTTG
ATCGGCTATGATCGGAACCCAGATTCTACAG
CAGGATGTCGTACAACGAAAGACATGGGTAA
AGTACAGTCTCCCTTGGCTGCTGCCCCACTGCC
TCTATAACACGCGGAGACTAAACTACGGAGTTC
GGTTCCATCGATACAGTGTGCTGCTGCCAATG
AATTTGCTGTTGGGTGTAATAATATGCCGA
TGAATCGGTTGAGGATGTAATCGTAGGAGATT
AGTATTCACAAGCTTAGAGGCTACGATATCTC
CCCTACAAGGTTGATGATGTTGGCTATCA
ATGGGGAACCGCTTATCTCTCTTTCTGAA
TATVAAGATAGGAACCCACTCCACTATCCAGGG
GCATTTCTGCAAGCTTAATAGTGGGGGCCAC
CAGAAGGCCCAAGCTTTGATGACCAATCACGTG
 \square CCGCGAATAATTTGAGAAAGGGTCC
TGAATGATTAAGCGGC
TATAAGGCTTCAAGGTAATTTGTAACCCCT
ATTAATTTGATTTGATTTGATTTGATTTGATTTG
TCTTTGATTTGATTTGATTTGATTTGATTTG
ACTTAACGACGTTGATTTGATTTGATTTGATTTG
TGTGATATGTTAAGAAATGATTTTACTCTGTA
TTGTCGAGGCTTCTCTCTTTGATTTGATTTG
ACAGCTTGTGACAGGAGTGTGATGATTTAA
CCATCTCGTGAAGTCTCTACAGATCTTGACAA
TGAAATGATGAGCACATGAGATGAGGGCTC
GAAGAACTGTTGTTGCTGACAGAGTATTTCCCT
GAAATGCTGCTGACTGCTGGGGATACCAAGTC
CGAACTTGAATTTAGCTCCCTGTAATGTTGGATG

Further analysis was focused on the
(β -component). Amplification of genomic
 β -component primer pair (Beta-01 and Beta-02)
fully produced about 1300 bp fragment. The
authors for instance Cotton Leaf curl virus (CLCV)
Mesta yellow vein mosaic virus (MYV) and
1359 bases and 1354 bases respectively. The
satellite DNA is longer than satellite DNA
from tomato leaf curl virus (ToLCV) which
632 bases (Figure 4).

Figure 4. Complete nucleotide sequence of
from TD-21 spanned 1347 bp. Sequence
leotide was primer sequence used in
reaction

GGACCCACTCCCTGGGGAAGATAAAGAG
TGGCCCTGGCATVAGGCCCATCATATTC
TTGGTGAATAAATGATCGGCTTAAAT
ACTTTTATTAAGTGAACGTTAAAG
ATCAAGTTTGGATACAAATAGAATAG
TACAAATGACCCCATTAATACATGAT
CCTAAATATATGCAATCATCTATCAAT
ICAGGATATATGCAATCATCTATCAAT
CTGCTTGAATCTCCAAATGGTAGATCC
ATGCTTTGAGCATATGGCATATGCTCT
GACTGTGAAGTGAATGGTATGATTA
GTGCCCTATGGATGAGGAACACTTT
AGTGTCTGACTTGGTGGAGACAAGT
GCACAAGAAATGATGATGAGCCCTTAT
GTGATGATGACGATGATCCCTTTGAT
TAGTGATGATGATGATGATGATGATGAT
GATGATGATGATGATGATGATGATGAT
TACGATGATGATGATGATGATGATGAT
TGCTCCCATGTTGTTGTTAAATGATG
CAGGATGATGTTGATGATGATGATGAT
TCAATGCTGCTGATGATGATGATGAT
GTTACTTATGCTGATGATGATGATGAT
CATGTAATTAAGAAAGAAAGTAAAT
AATGATCCACCATGATTAAGAAAGAA
TAGAAGGCAAAACCATAAACCTAA
AGAAGAACTAGATGAGAGATATAT
AGAAATGGAGGCAAGCAAAAAGTCA
TCTAATTCACAAAACCAAAAGTCA
CCTTAAATCTGCAAGTCCGACTTCC
TATTACTGATTAACACAGGTAATTA
TGCTATTAACCCCTGTTAAATGATTA
ATWAGTCCCAATAGGTAATGTTAA
TCGGATACCAATTTGAGACAGAAAG
AATAAAGAACCAAAAATACCTTAA
TCTATTAGGGCGGTGGAGTCTGAA
AGACCTTCTCTCCAAAACCTACCG
GATACAGGCTGATCCCGCATCAATTT
GCCCGGGG

... virus: karakteristik isolat Cabai dan ... Cucumber Mosaic Virus dan Chili ... Institut Pertanian Bogor, Bogor.

... Kuning Cabai. Disertasi Seko- ... DNA Virus Penyebab Pe- ... Karakterisasi Biologi, Serologi ... 5463-5467

... of the National Academy of Sciences ... DNA se- ... Coullson, A.R., & Coullson, S. ... own genome. J. Theor. ... deriving resistance ... 1985. The concept ... Johnson, 1985. Plant Dis. 77:340-347.

... Whicfly ... of Degenerate Primers in the ... D.R. Russell, and D.P. ... 1285-1290.

... two tomato infection gem- ... Demisia tabaci (Homop- ... Polymerase chain reaction ... Wymann, M.K., Nakhta, ... Harbor Laboratory ... Laboratory Manual, United ... and J. Sambrook. 1989.

... 16 - 18 September 1999 ; ... Fitopatologi Indone- ... Kongres Nasional XV dan ... virus gemini pada cabai ... dalam *polymerase chain* ... N. Aidawati. 1999. Peng- ... 336-342.

... Hidayat, and R.T. Martinez. ... (1): (1983) 19-21.

... Wood, J. and Hicks, J.B., *Plant ...* 343-354.

... Muthi and S. K. Mukherjee.

... and Development Agency of Agri- ... of this research. This ... via KKP3T Programme of fiscal

... of satellite DNA showed 1347 ... DNA-B ... genome containing only DNA-